

Hemolytic Uremic Syndrome

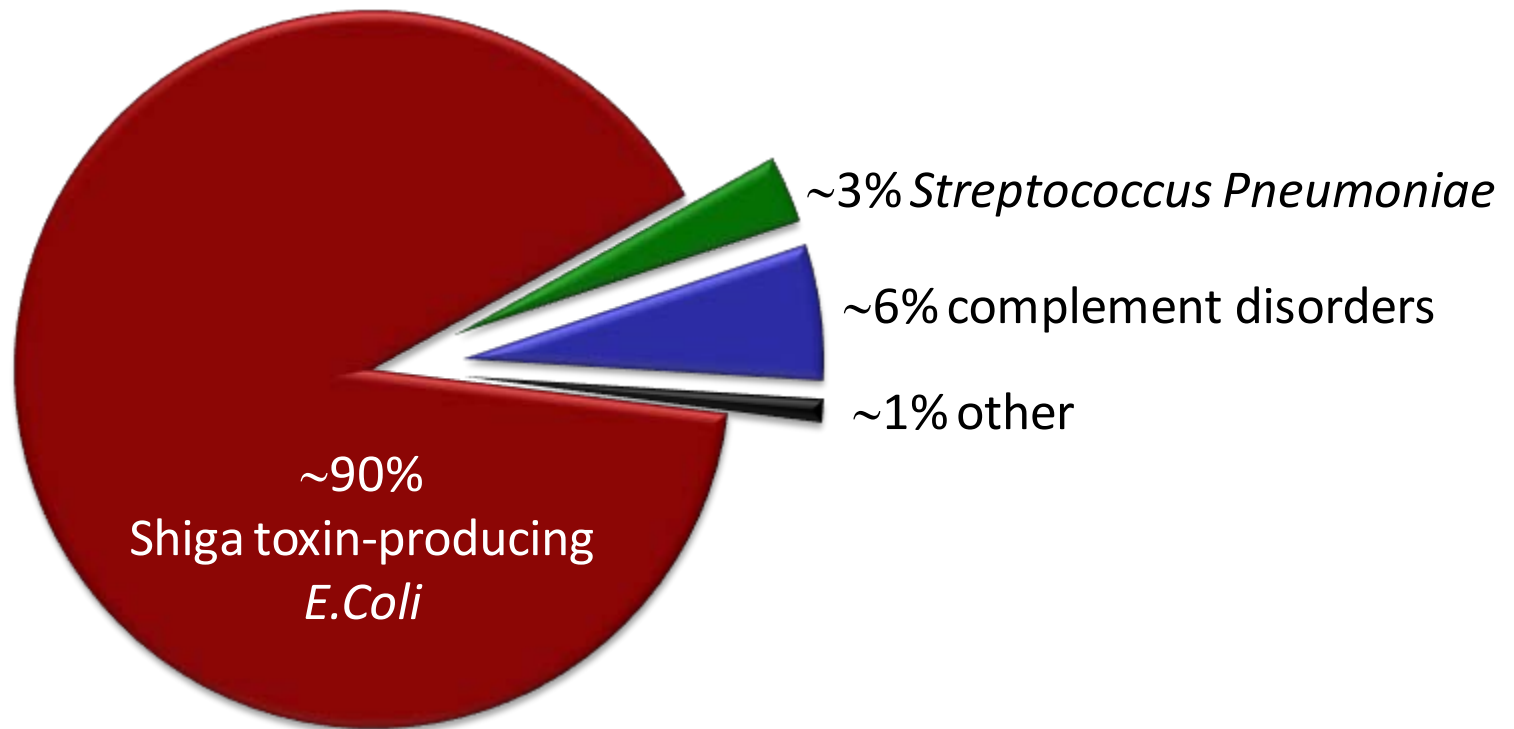
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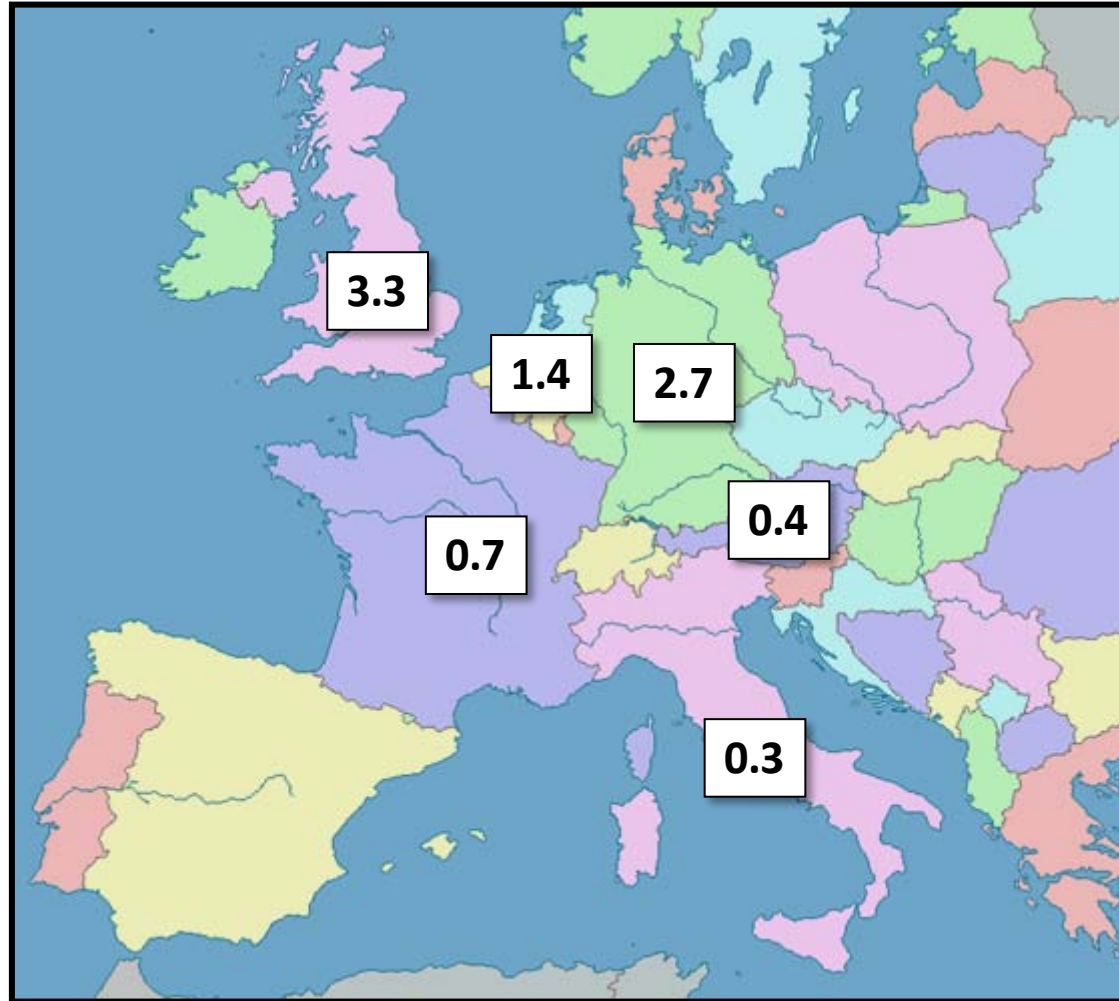


Hemolytic Uremic Syndrome (HUS)

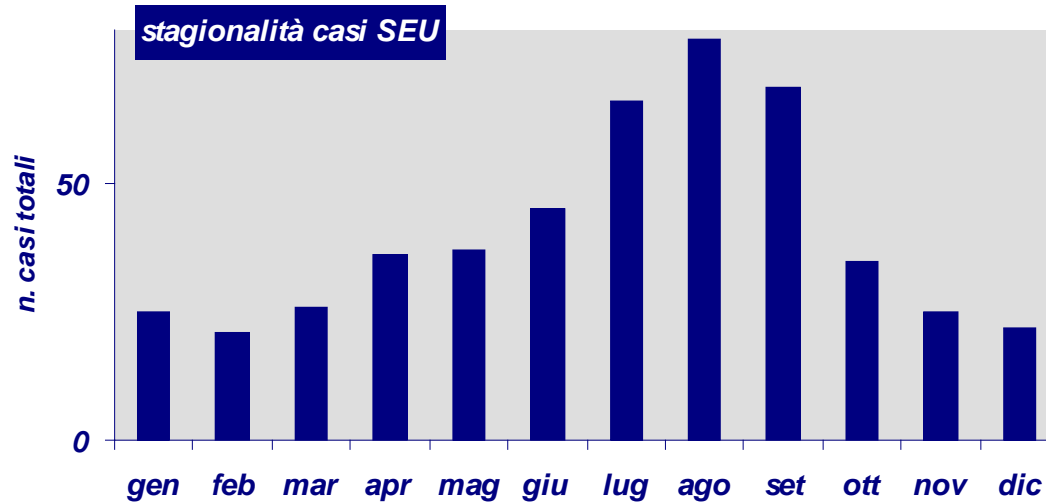
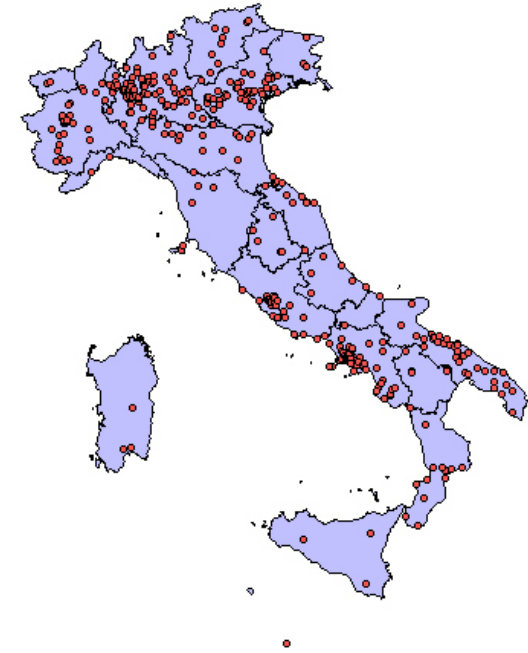
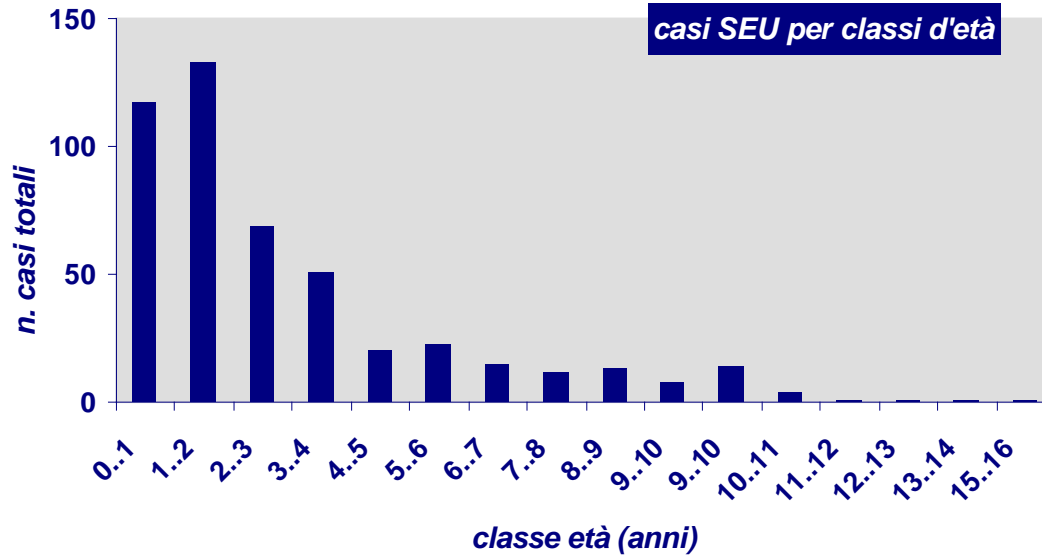
microangiopathic hemolytic anemia
thrombocytopenia
acute nephropathy



HUS: incidence (cases / 100,000 x year)



Italian Registry of Hemolytic Uremic Syndrome



N=481

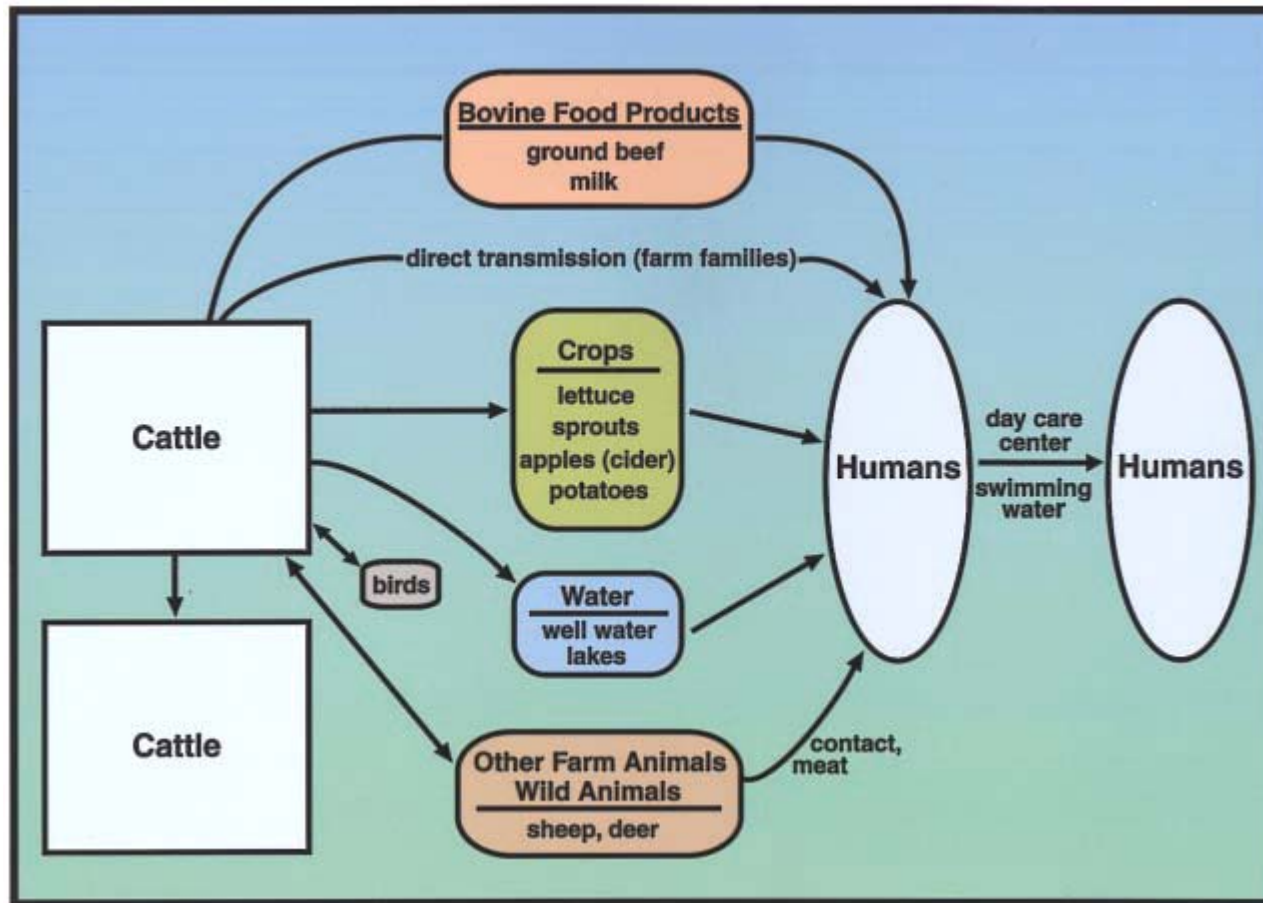
53:47 M:F

Median age : 23 months

< 5 years = 80% of cases

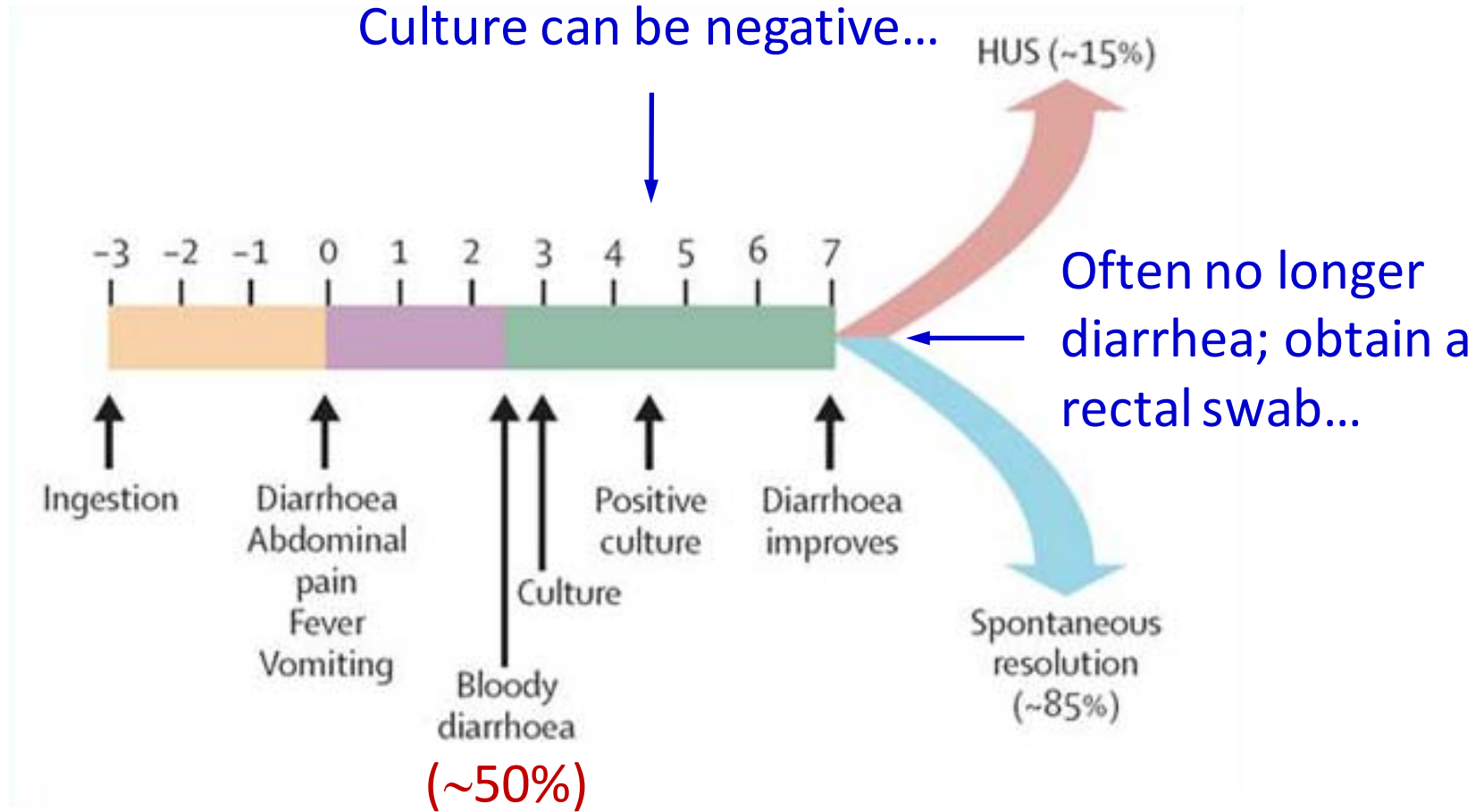
< 2 years = 41% of cases

Model of transmission of *E. Coli* O157:H7 from cattle to humans



E. coli O157:H7 Infection: time course

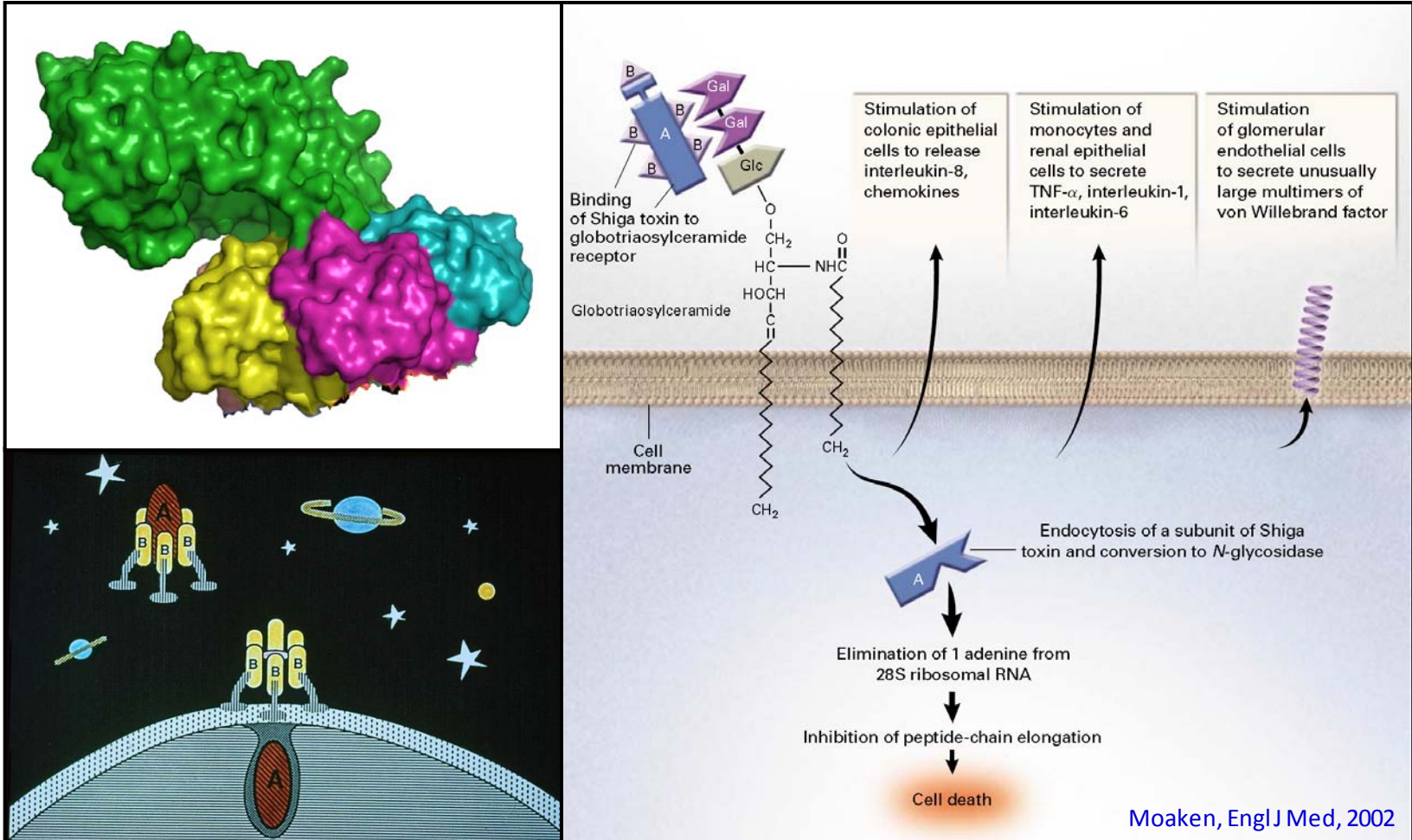
Often antibiotics
Culture can be negative...



Shiga toxins

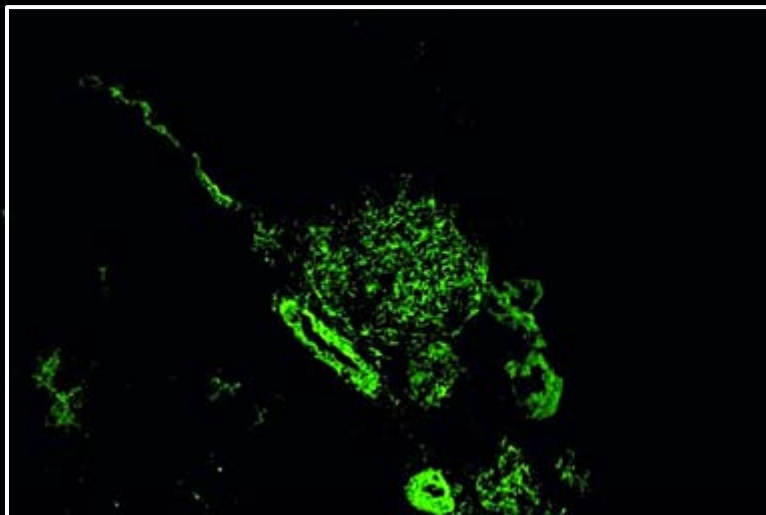
- ❑ **ST1**: identical to toxin of Shigella
- ❑ **ST2**: 56% homology; 30X more potent
- ❑ E.coli O157:H7 ~100% ST2; 25-80%ST1
- ❑ Non O157 STEC
 - O111:H8 (Colitis)
 - O26:H11
 - O103:H2 (UTI)
 - O103:H3
 - other
- ❑ Single A subunit, 5 B subunits

Shiga toxins

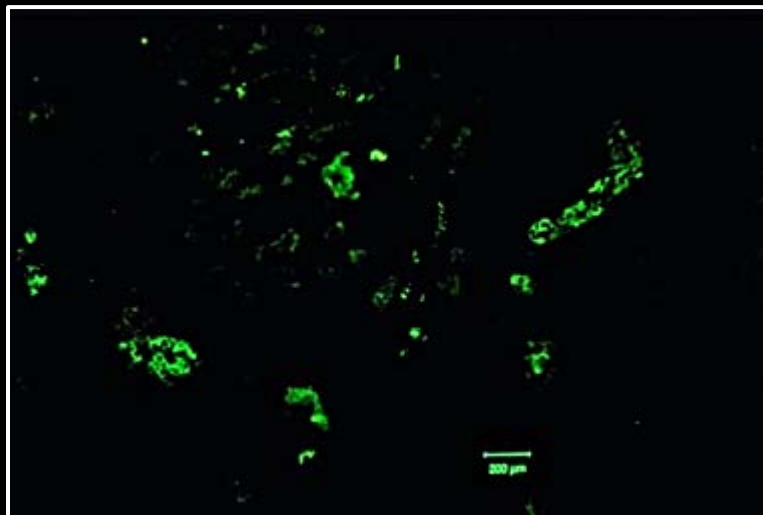


Gb3 expression in human kidneys

Pediatric

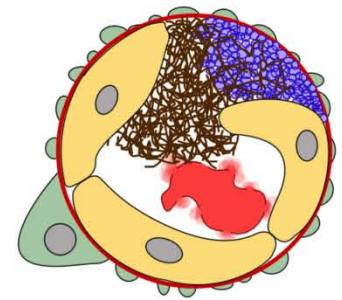
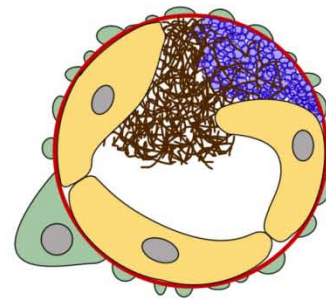
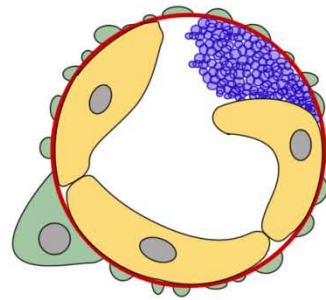
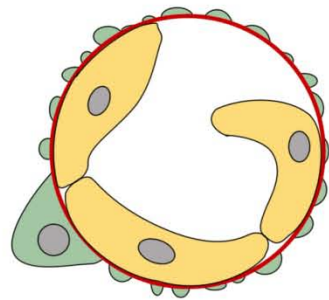
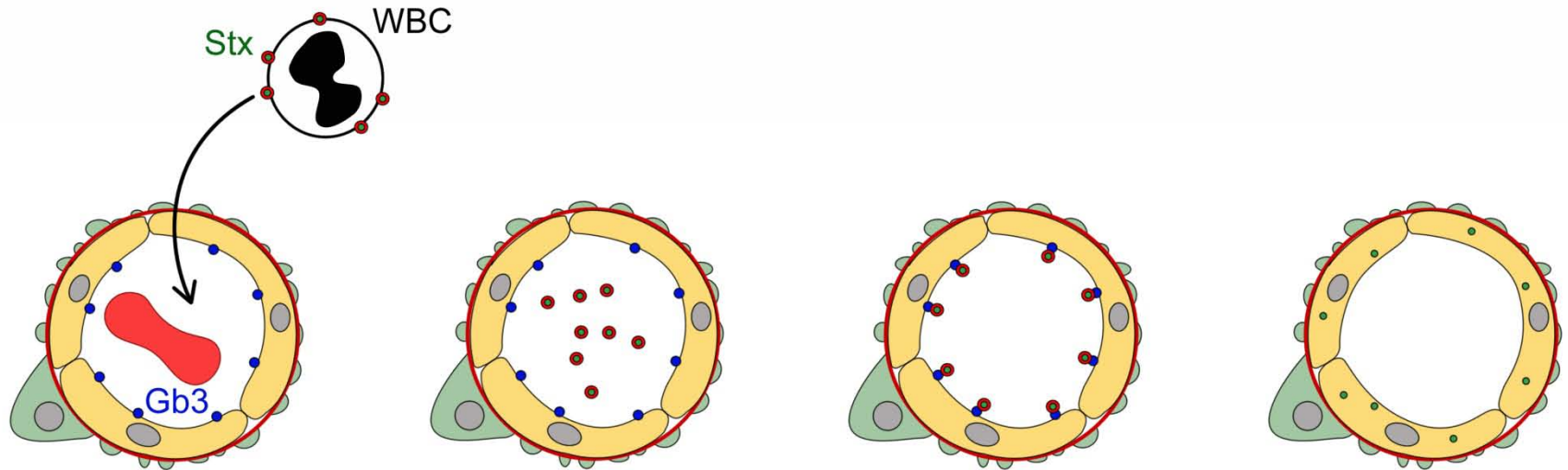


Adult



Lingwood CA, 2000

STEC-HUS



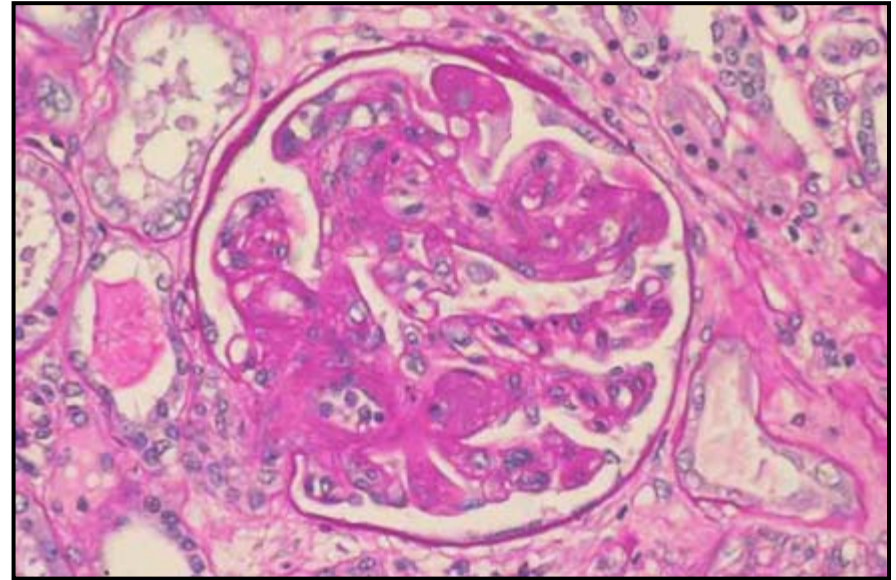
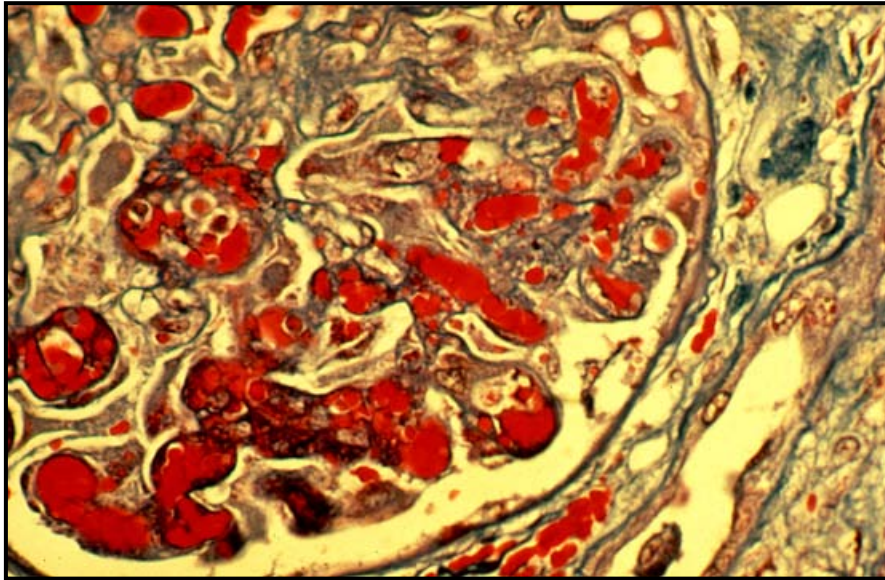
cell swelling and death
endothelial cell detachment
basement membrane exposure

platelet activation

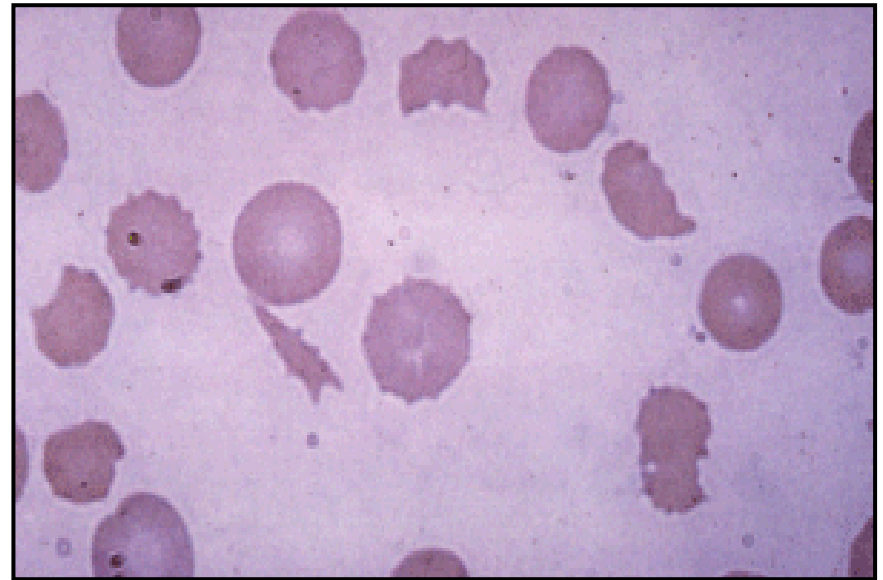
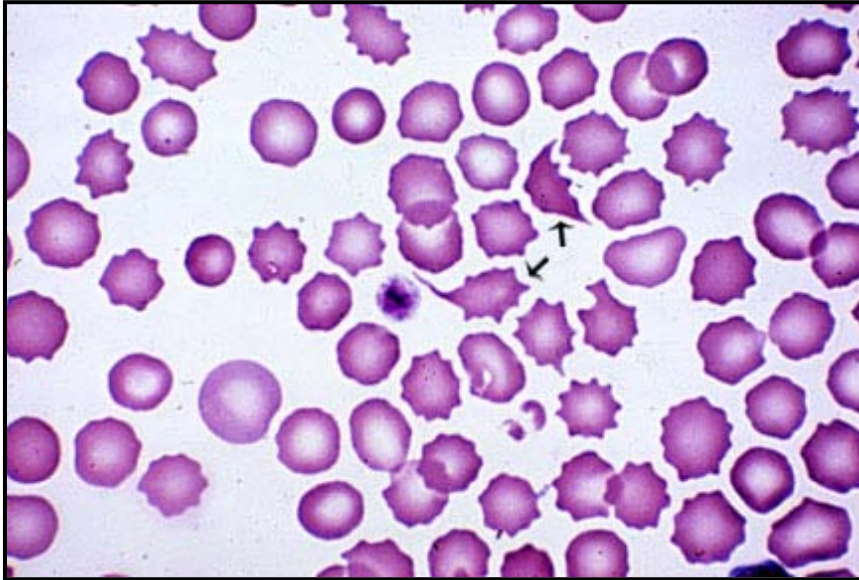
fibrin deposition

RBC destruction

Renal pathology: acute

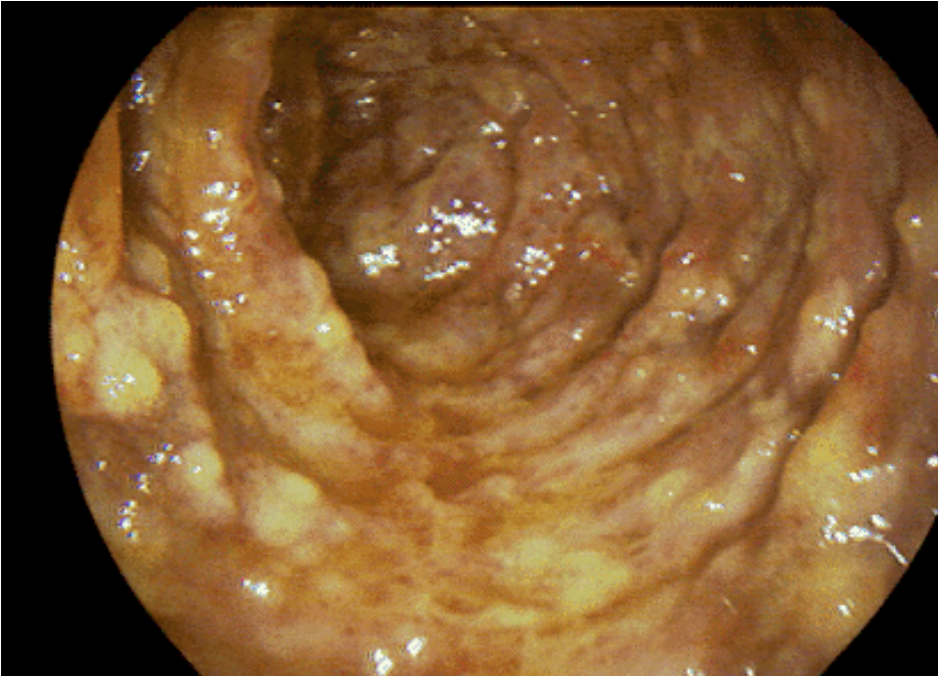


Microangiopathic hemolysis



- Poor correlation with the degree of ARF
- High LDH

Gastrointestinal involvement



- Colitis, perforation (2%)
- Prolapse (10%)
- Pancreatitis (20%)
- Diabetes mellitus (8%)
- Elevated LFT's (40%)

CNS involvement

❑ Symptoms

Seizures

Coma

Stroke

Cortical Blindness

❑ Etiology

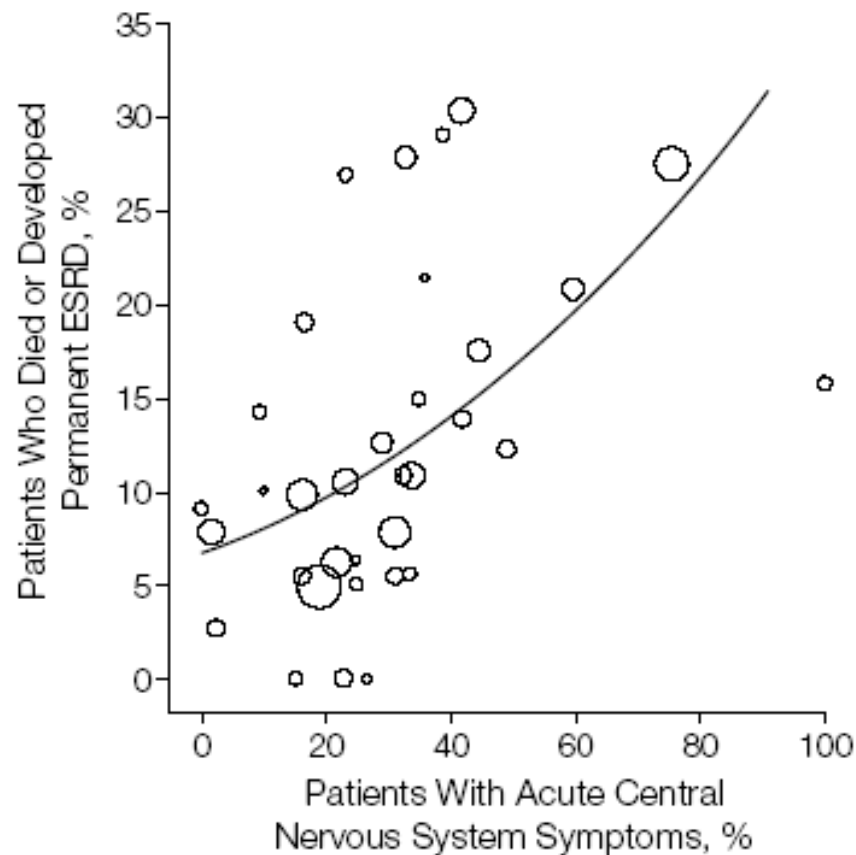
metabolic (high BUN, low Na⁺, low Ca⁺⁺)

HTN

TMA

CNS involvement

Figure 2. Studies With a Higher Proportion of Patients With Central Nervous System Symptoms (Coma, Seizures, or Stroke)



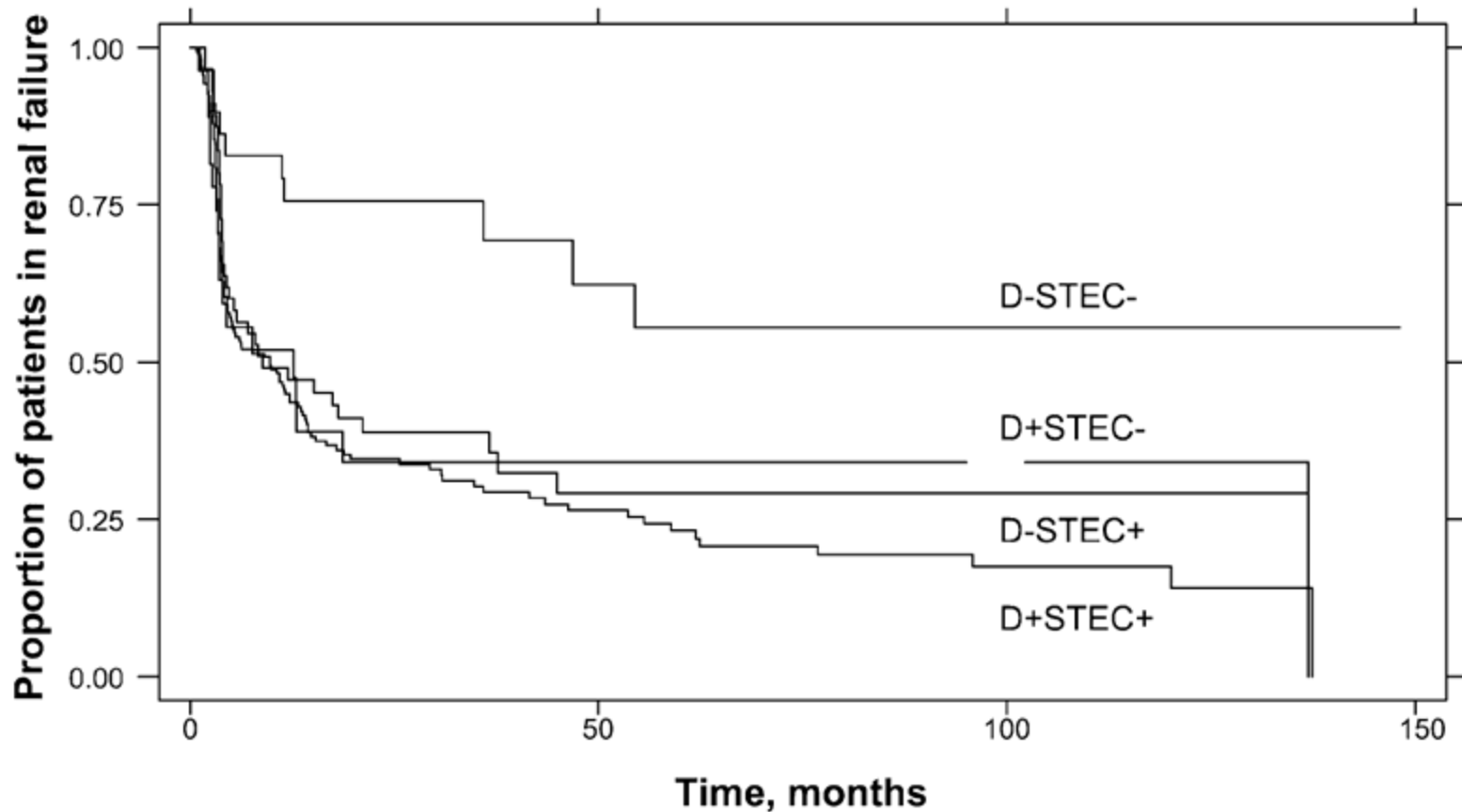
Reported risk factors for poor outcome

Reference	N	Oliguria	CNS	age	↑ WBC	D -
Trompeter et al 1983	72			older		+
Loirat et al 1984	67	> 8 d		> 3 y		-
Walters et al 1989	79				+	
Coad et al 1991	74				+	
Siegler et al 1991-96	118	>15 d		< 2 y	+	±
Lopez et al 1992	509				-	+
Robson et al 1993	71	> 10 d				
Kelles et al 1994	95	no	+	no		-
Tönshoff et al 1994	89	> 14 d	+	no	-	+
Renaud et al 1995	42					+
Spizziri et al 1997	312	> 7 d		no	-	
Huseman et al 1999	149	> 7 d	-			
Gianviti et al 2003	387		-	no	-	+

Supportive management of STEC-HUS

- Fluid resuscitate if dehydrated
- Avoid volume overload
- Treat hypertension
- Transfuse if Hb < 6-7 or earlier if symptomatic
- Avoid platelet transfusions
- Dialysis as needed
- Plasmapheresis ???
- Nutrition (TPN)

Italian Surveillance Study 1988-2002



Group	No. patients	Total proportion recovered (95%CI)	No. deceased patients
D+STECS+	157	75.8% (68.2–82.1)	1
D-STECS+	27	66.7% (46.0–82.8)	0
D+STECS-	55	65.4% (51.3–77.4)	1
D-STECS-	29	34.5% (18.6–54.3)	3

Atypical HUS

❑ Other pathogens

S. Pneumoniae

HIV, Q fever, CMV, Staphylococcus, Hantavirus

❑ Drugs

cyclosporine, tacrolimus

bleomycin, cisplatin, mitomycin

❑ Underlying genetic defects

complement factors deficiency

cobalamin-C defect (methylmalonic aciduria and homocystinuria)

Streptococcus Pneumoniae-associated HUS

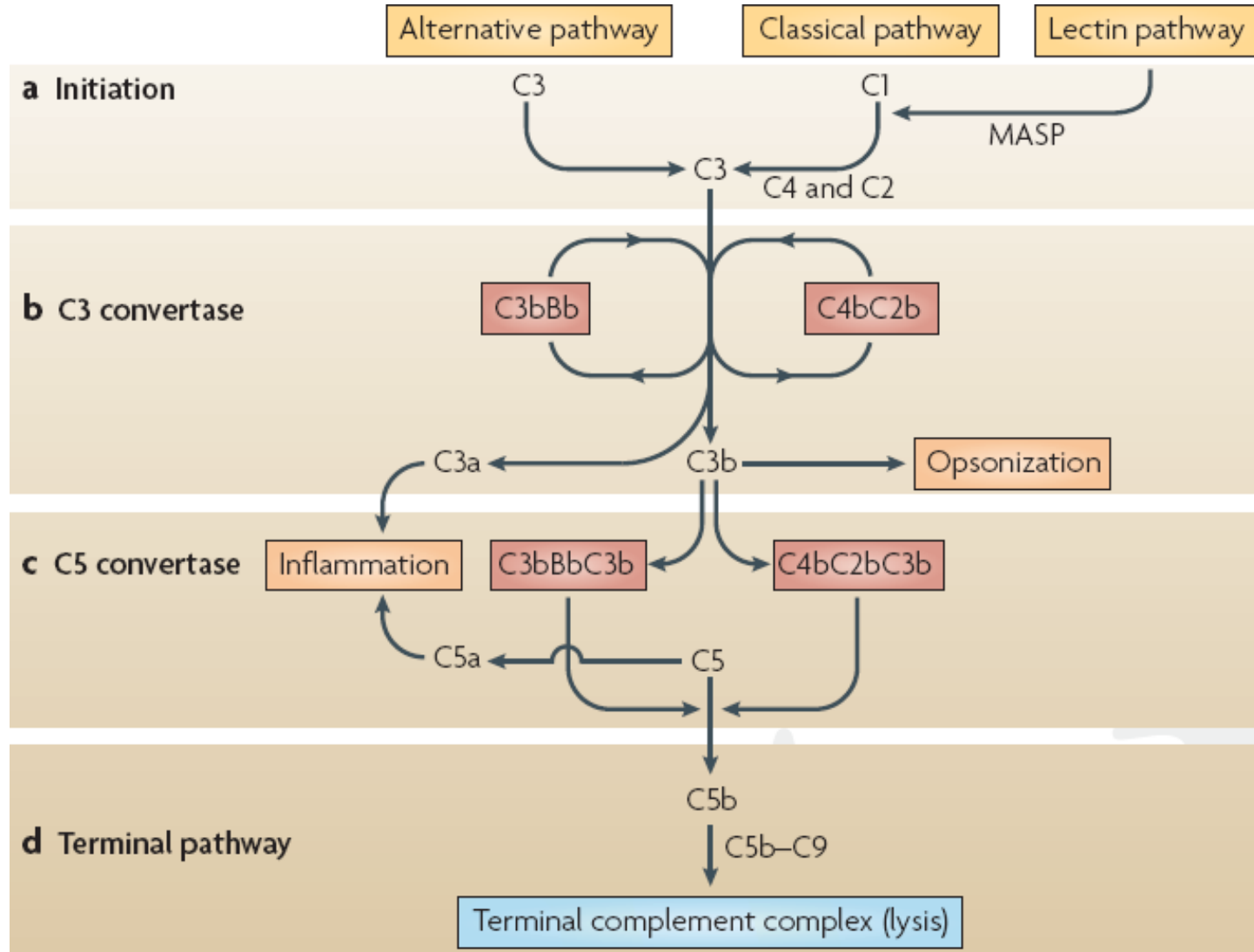
- ❑ Nearly always invasive disease
 - ± 60% empyema
 - ± 30% meningitis

- ❑ Overall mortality: 13% (30% if meningitis)

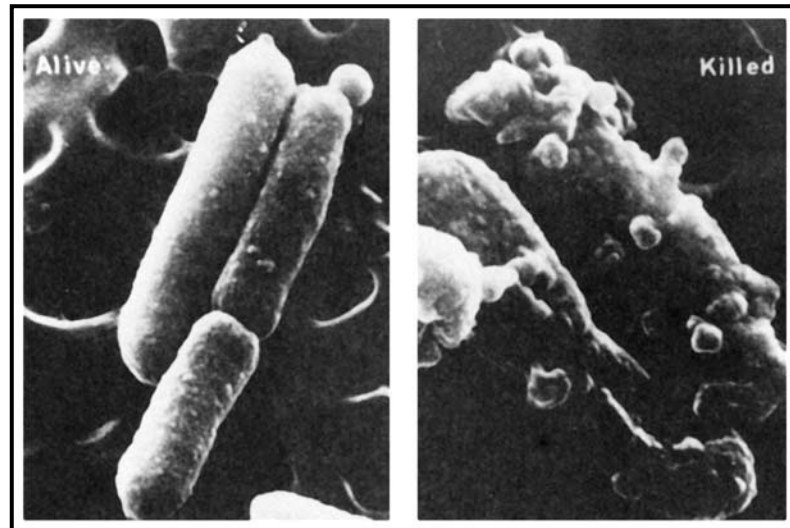
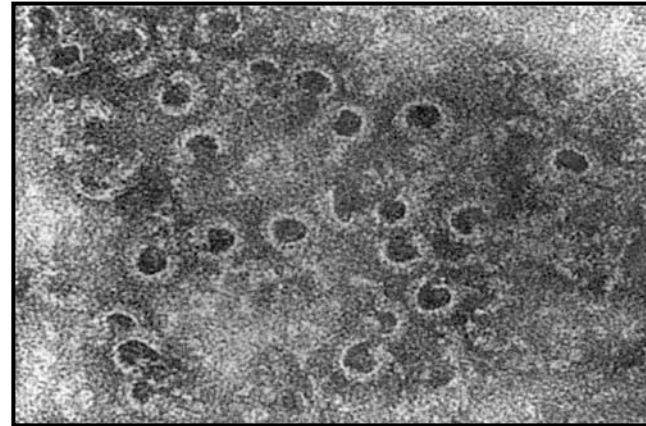
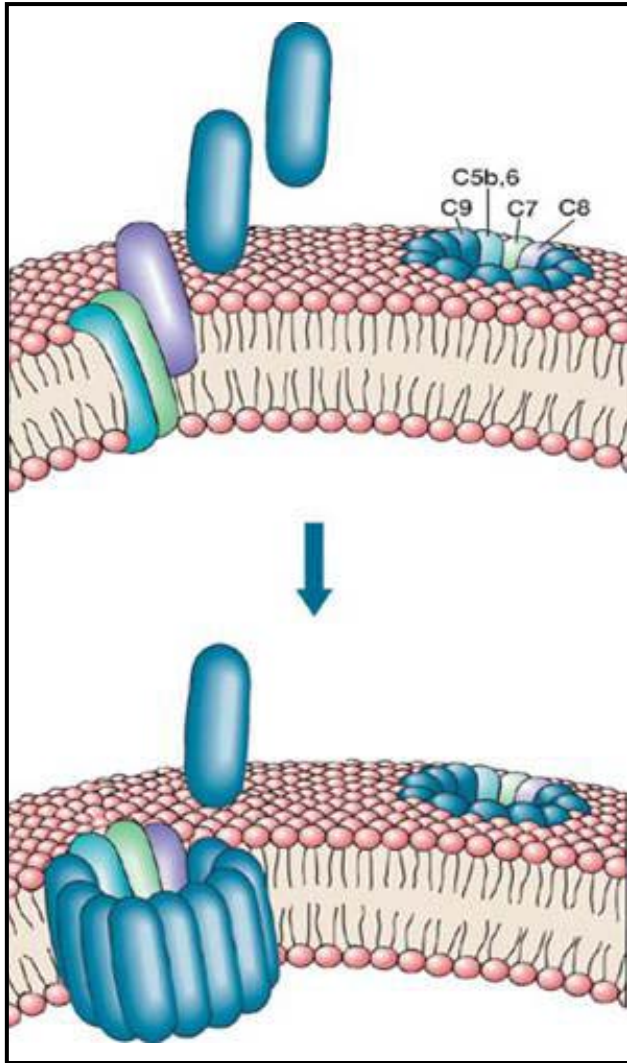
- ❑ Acute phase more severe than D+HUS
 - more dialysis (84%)
 - more transfusions (98%)

- ❑ Long term renal sequelae similar to D+HUS
 - protU and/or CRF 20-25%

A simplified view of the complement system



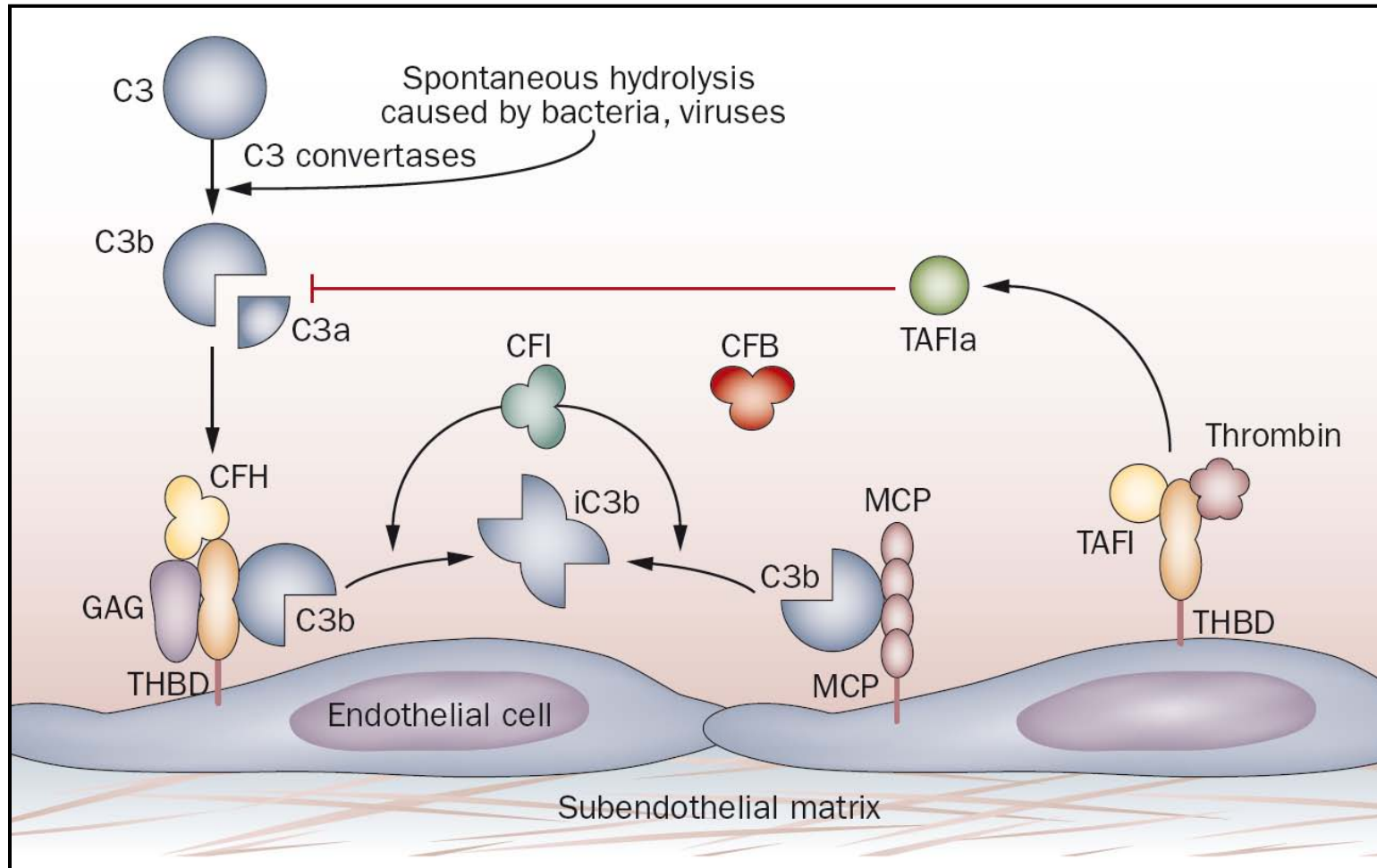
Membrane Attack Complex (MAC)



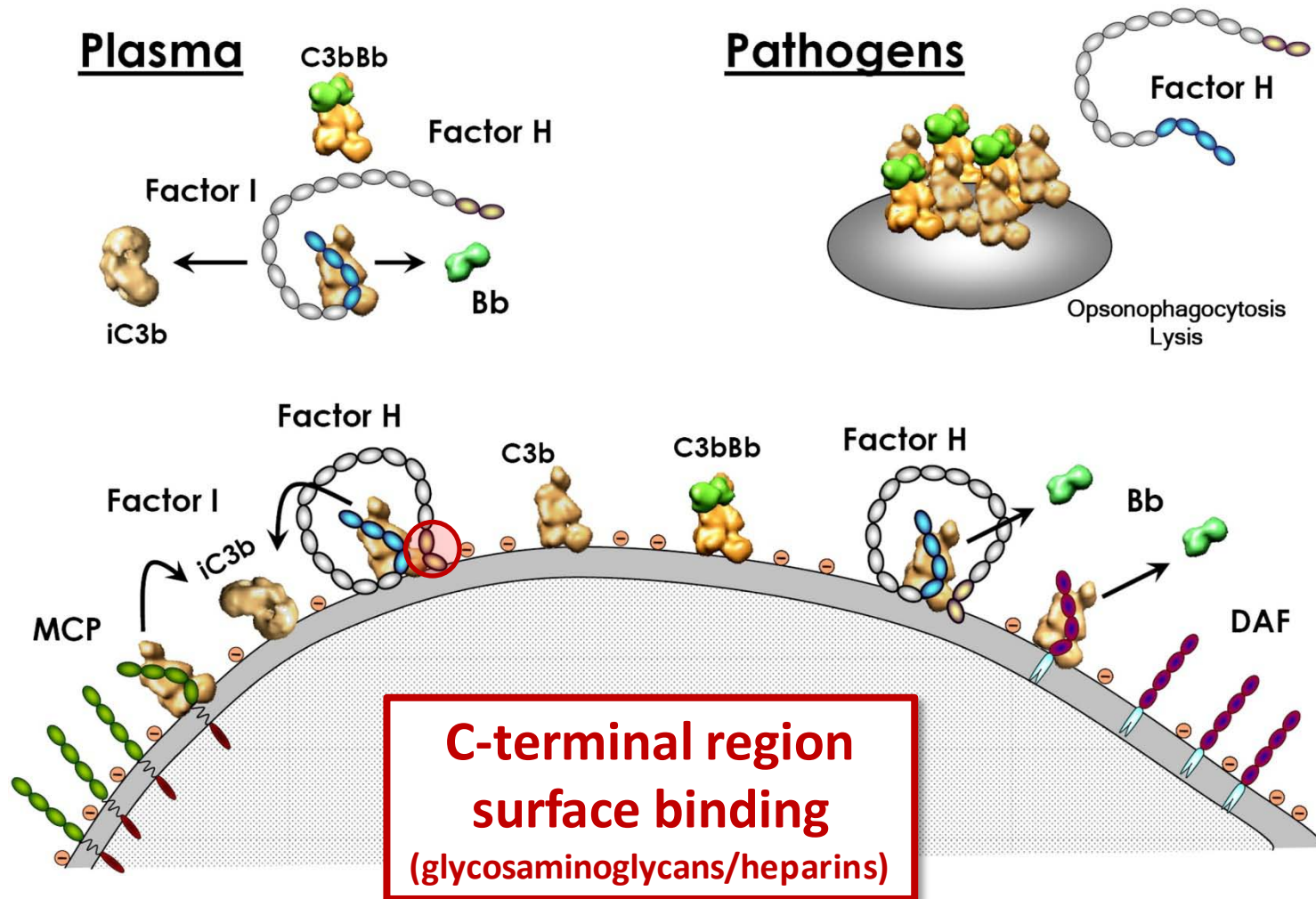
The area of complement....

- ❑ Erroneous activation or insufficient regulation of the complement cascade may lead to an attack by the immune system against self-tissue.
- ❑ Many autoimmune, inflammatory and ischemia/reperfusion injury-related diseases are connected with complement.
- ❑ Some of pathological conditions are localized to specific tissues, but many are systemic.

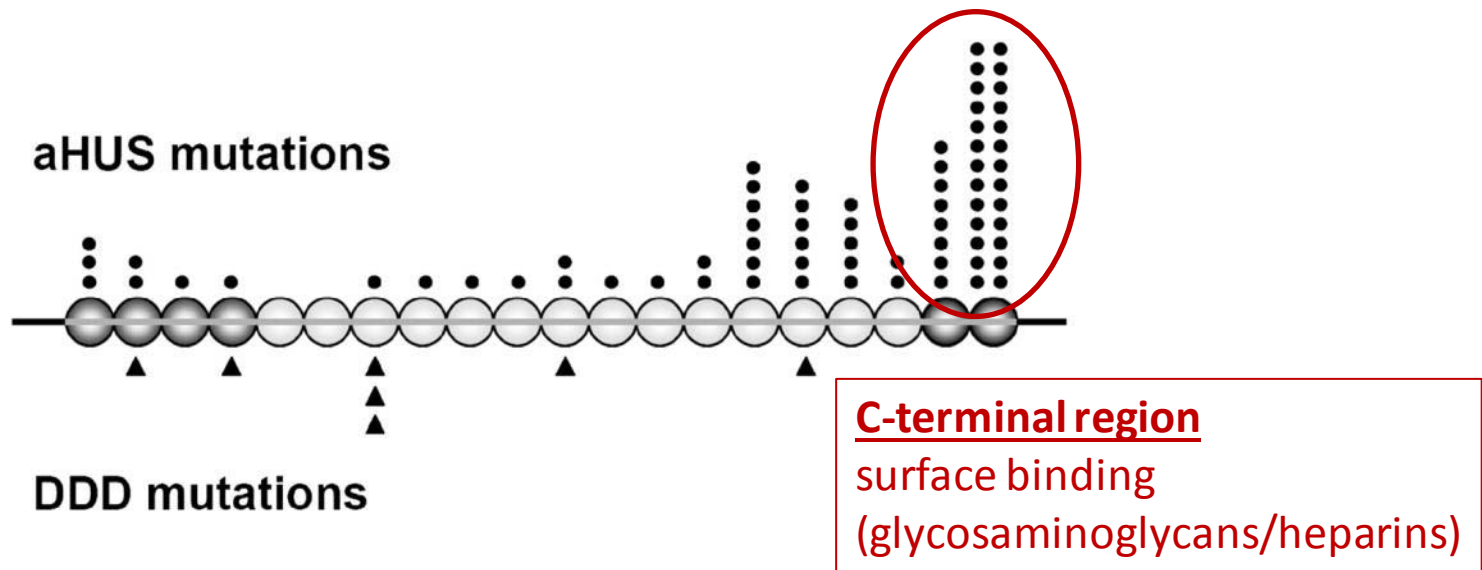
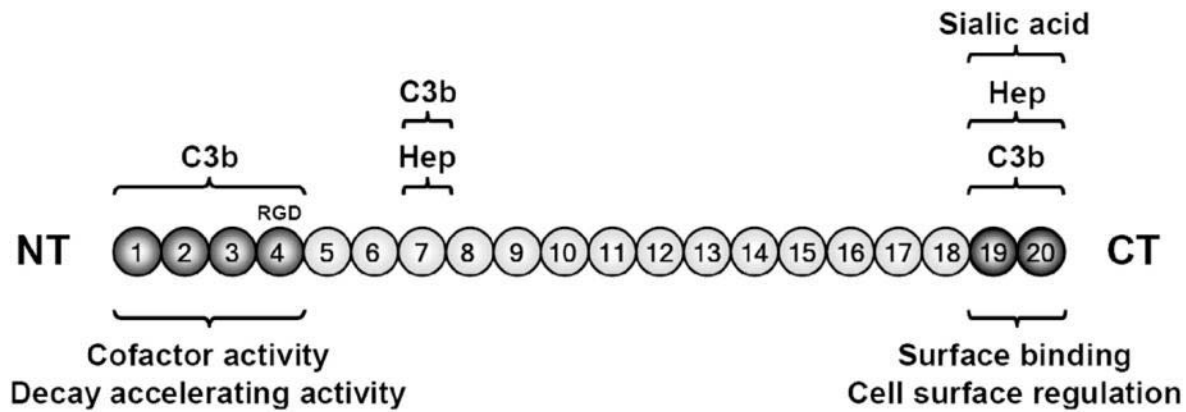
Endothelial cells with functional complement regulation



The AP can be activated on cell surfaces or in the fluid-phase

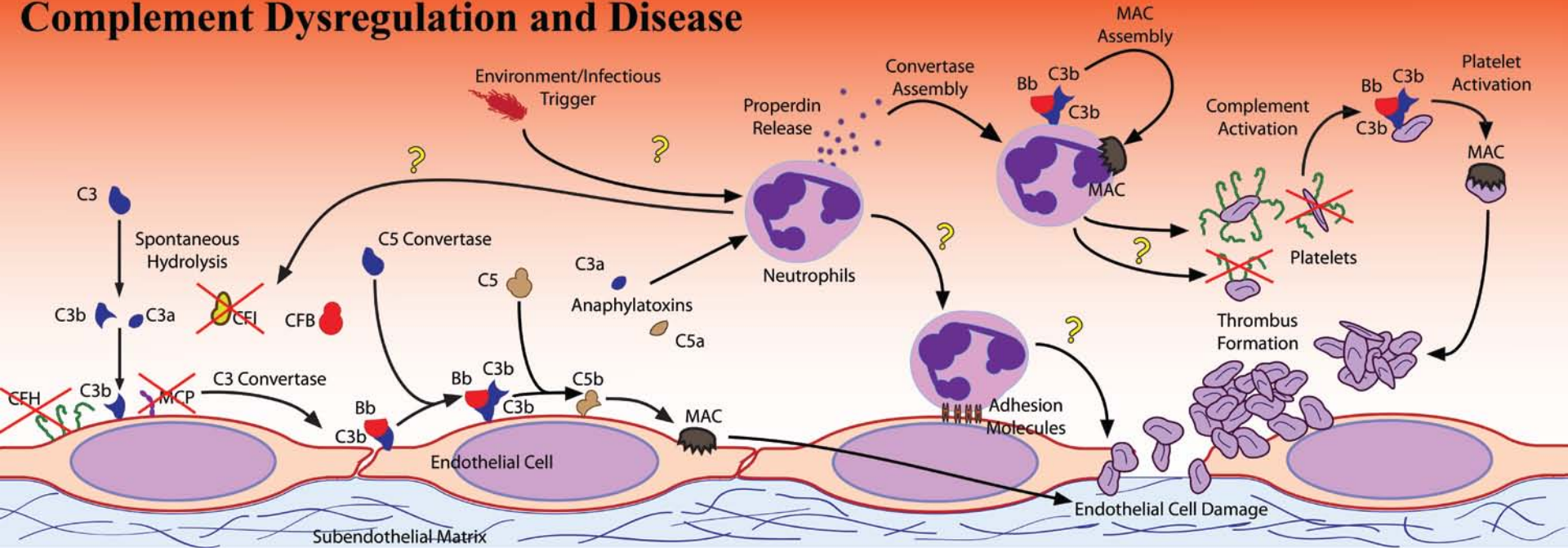


CFH domain mutations in aHUS and DDD



Thrombotic microangiopathy

Complement Dysregulation and Disease



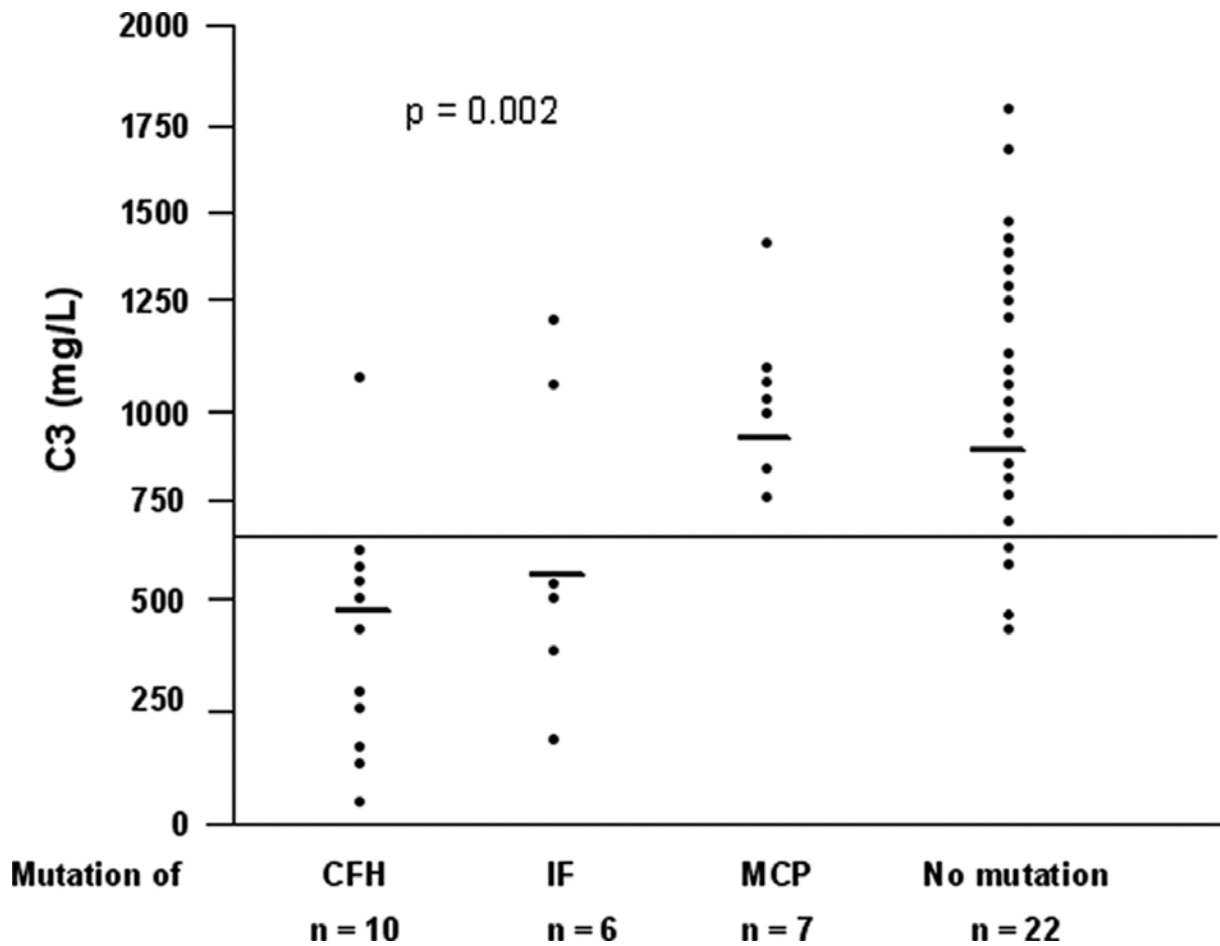
aHUS related complement mutations

- Loss of function mutations:
 - Factor H (CFH)
 - Factor I (CFI)
 - Membrane cofactor protein (MCP; CD46)
 - Thrombomodulin (THBD; CD141)
- Gain of function mutations:
 - Factor B (CFB)
 - C3
- Autoantibodies:
 - CFH autoantibodies \pm CFHR3-CFHR1 deletion (DEAP-HUS)

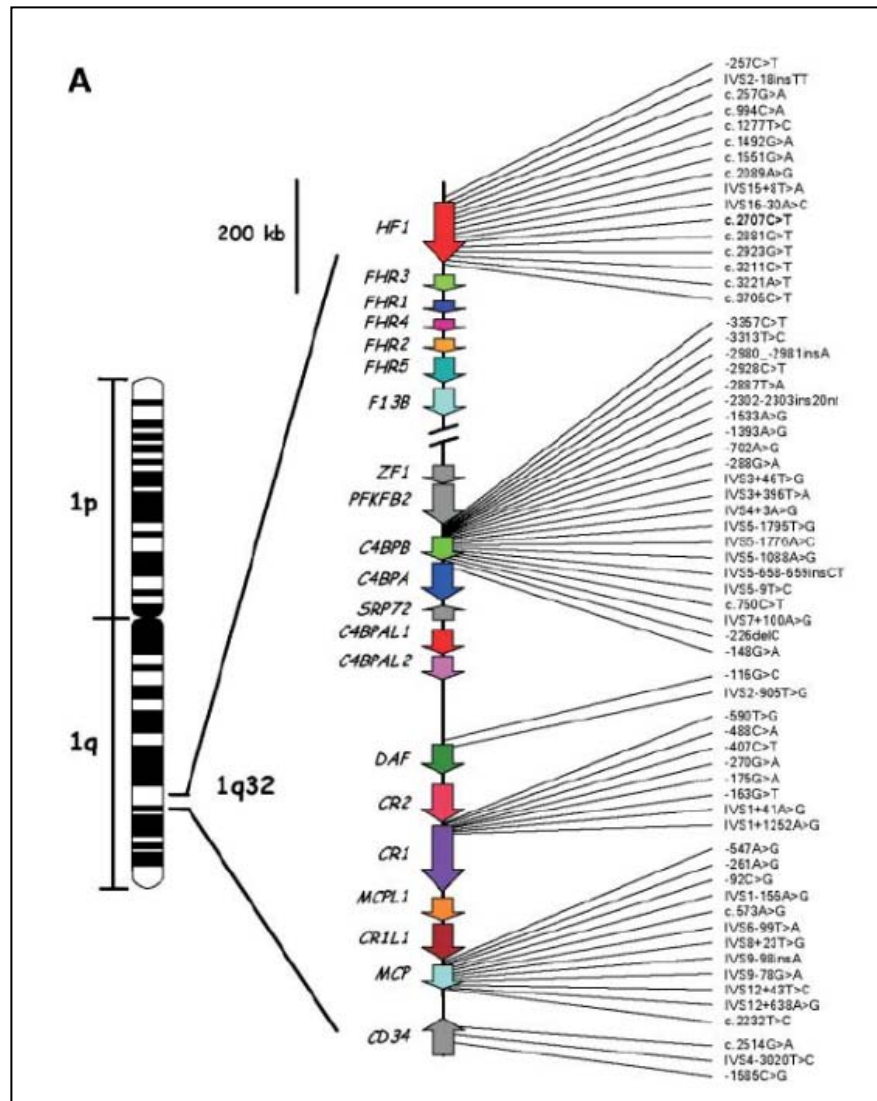
Genotype – phenotype correlation

Gene	Frequency	Response to plasma therapy [Remission]	Long-term outcome [Death or ESKD]	Outcome after kidney transplantation [Recurrence]
CFH	20-30%	60%	70-80%	80-90%
CFH Abs	6(-10)%	70-80% (PE + IS)	30-40% ESKD	20%
MCP	10-15%	Not indicated	<20%	15-20%
CFI	4-10%	30-40%	60-70%	70-80%
CFB	1-2%	30%	70%	Recurrence in 1 case
C3	5-10%	40-50%	60%	40-50%
THBD	5%	60%	60%	Recurrence in 1 case

C3 levels by mutation



Risk haplotypes in aHUS

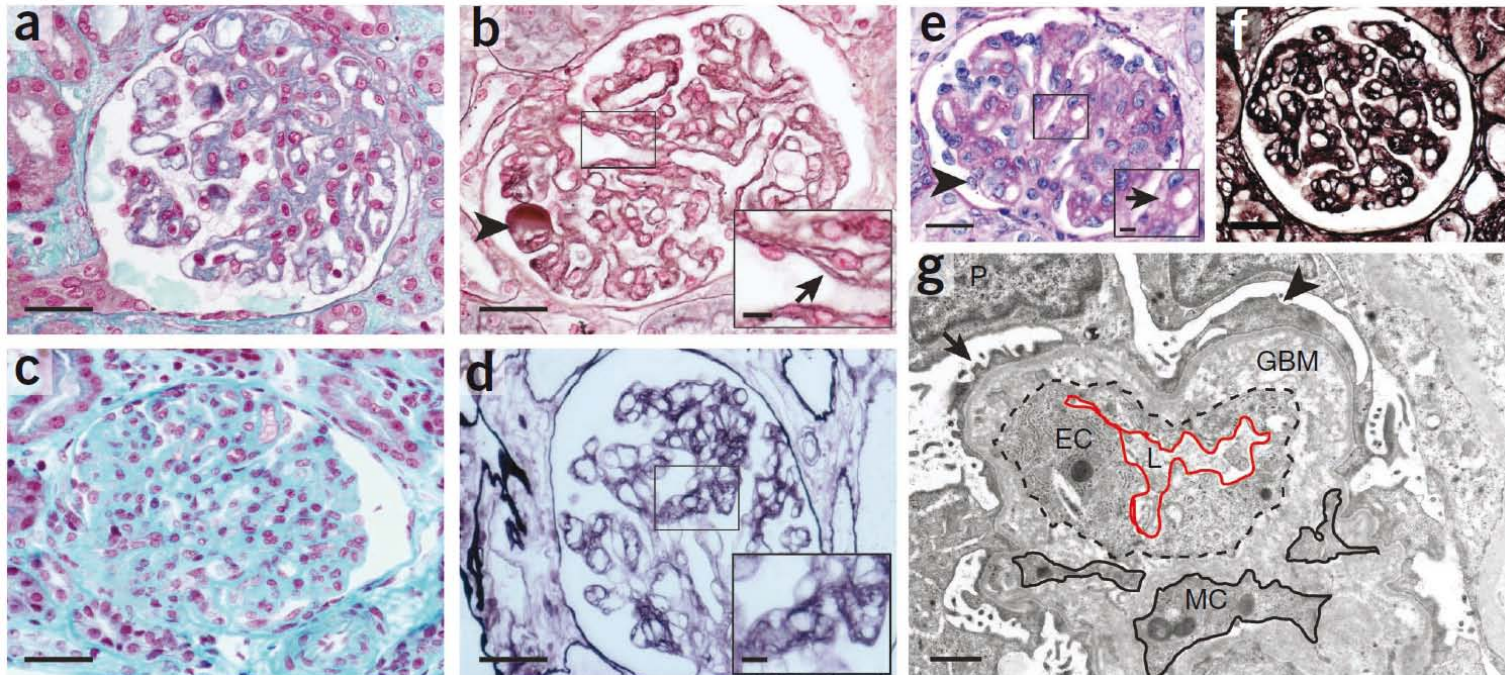


DGKE (diacylglycerol kinase ϵ) mutations

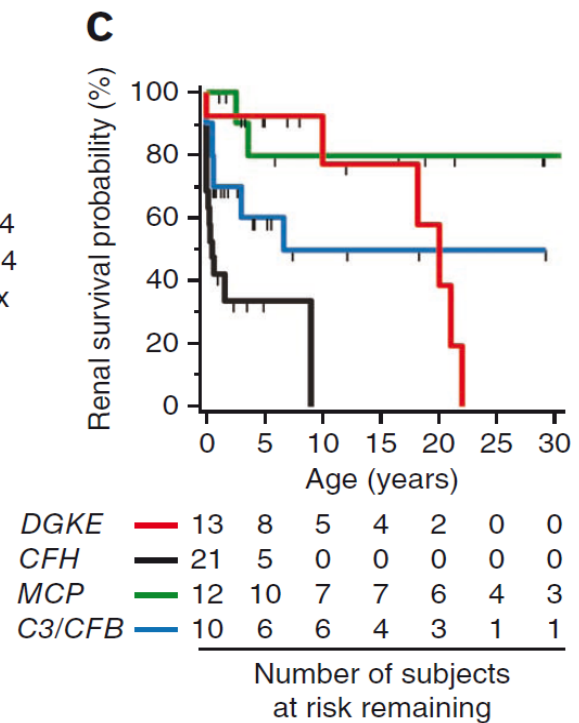
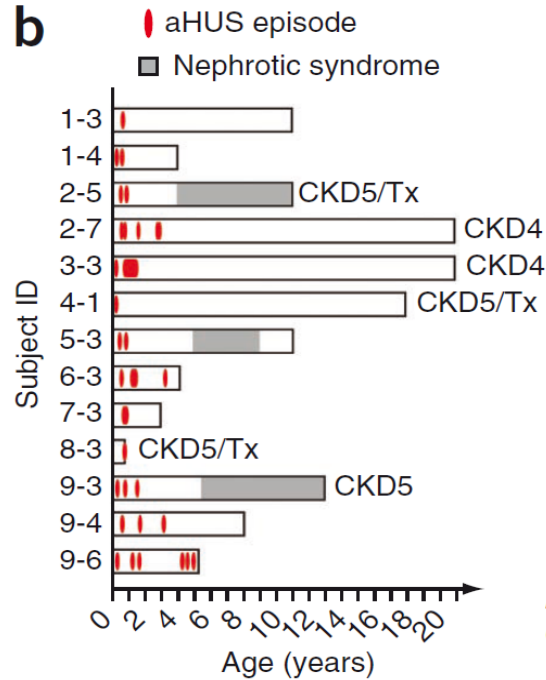
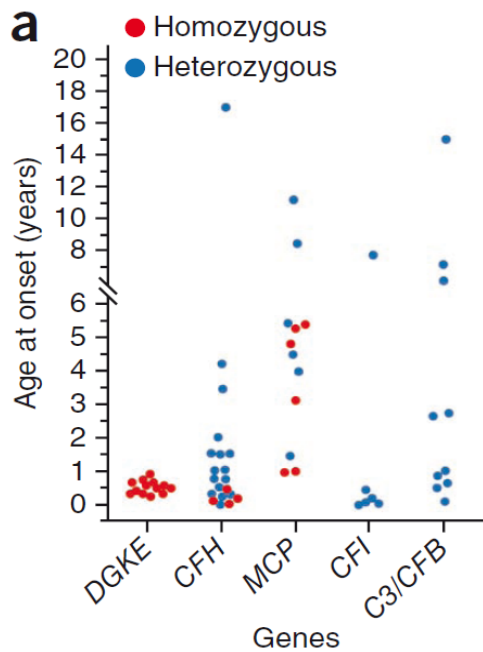
Recessive mutations in *DGKE* cause atypical hemolytic-uremic syndrome

nature
genetics

Mathieu Lemaire^{1,2,24}, Véronique Frémeaux-Bacchi^{3,4,24}, Franz Schaefer^{5,6}, Murim Choi^{1,2,7}, Wai Ho Tang⁸, Moglie Le Quintrec⁴, Fadi Fakhouri⁹, Sophie Taque¹⁰, François Nobili¹¹, Frank Martinez¹², Weizhen Ji^{1,2}, John D Overton^{1,7}, Shrikant M Mane^{1,7}, Gudrun Nürnberg¹³, Janine Altmüller¹³, Holger Thiele¹³, Denis Morin¹⁴, Georges Deschenes¹⁵, Véronique Baudouin¹⁵, Brigitte Llanas¹⁶, Laure Collard¹⁷, Mohammed A Majid¹⁸, Eva Simkova¹⁸, Peter Nürnberg^{13,19,20}, Nathalie Rioux-Leclerc²¹, Gilbert W Moeckel²², Marie Claire Gubler²³, John Hwa⁸, Chantal Loirat¹⁵ & Richard P Lifton^{1,2,7}



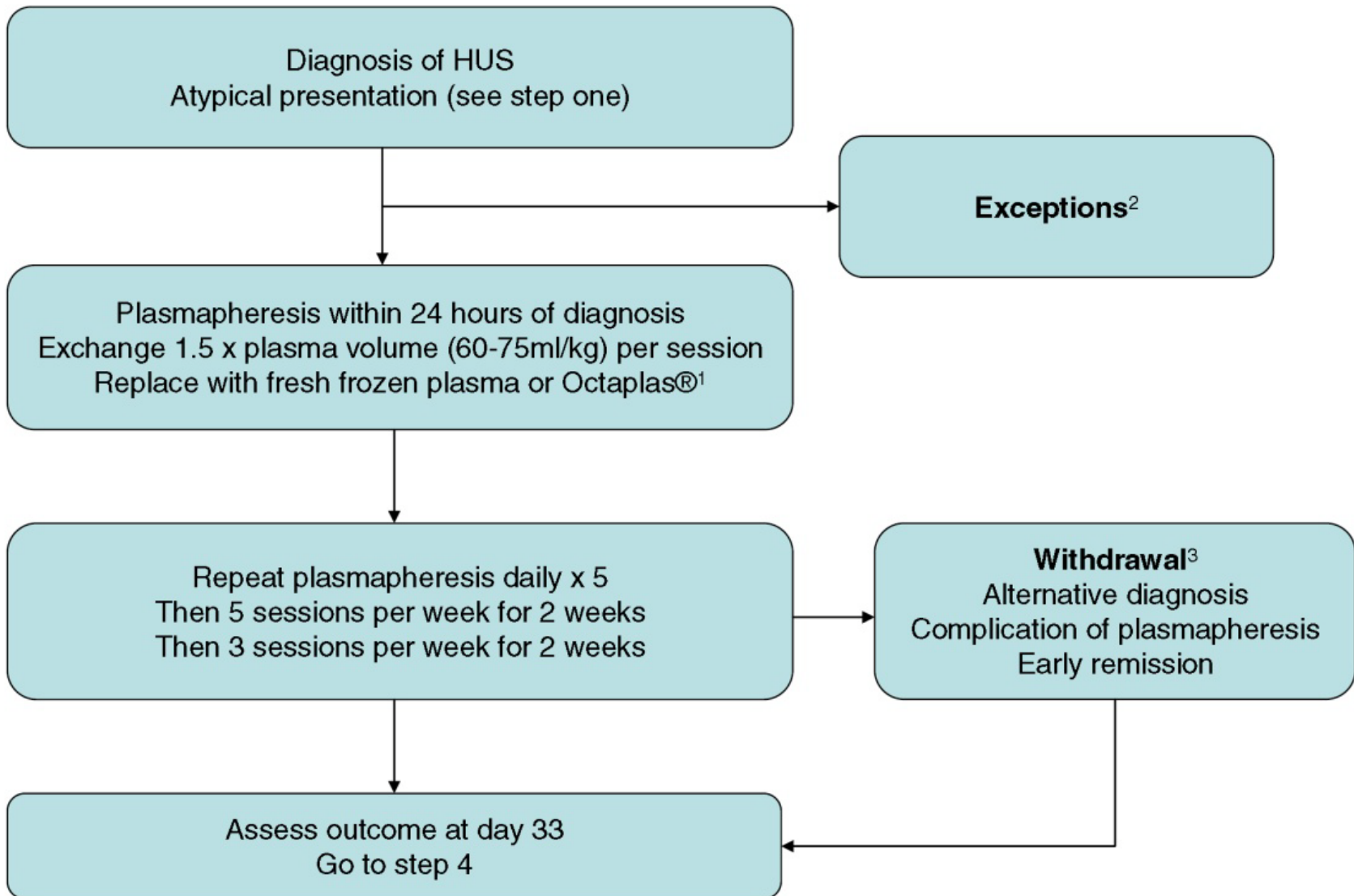
DGKE (diacylglycerol kinase ϵ) mutations



DGKE (diacylglycerol kinase ϵ) mutations

- ❑ DGKE is expressed in endothelium, platelets and podocytes
- ❑ DGKE inactivates arachidonic acid signaling through diacylglycerols (DAG) (activates thrombosis)
- ❑ First gene implicated in aHUS that is **not** an integral component of the complement cascade
- ❑ Symptoms: hypertension, microhematuria and proteinuria and unique finding of nephrotic syndrome
- ❑ Relapse of aHUS in subjects while receiving anti-C5
- ❑ Renal transplantation could be efficacious and safe

Plasmapheresis for aHUS



Liver-kidney transplantation

SPECIAL ARTICLE

www.jasn.org

J Am Soc Nephrol 20: 940–949, 2009.

Liver-Kidney Transplantation to Cure Atypical Hemolytic Uremic Syndrome

Jeffrey M. Saland, Piero Ruggenti, Giuseppe Remuzzi, and the Consensus Study Group

Eligibility for combined liver and kidney transplant or liver transplant alone

CFH or *CFI* mutation with the exceptions listed in Table 7

Less than 10% normal *CFH* levels in plasma

Patients who have identified mutations of genes encoding and have aHUS recurrence after isolated kidney transplantation or

have a family member who had the same mutation and had aHUS recurrence after isolated kidney transplantation

HUS recurrence after isolated kidney transplantation in patients with identified mutations of genes that may have both hepatic and nonhepatic sites of expression and protein synthesis (see Table 7)

Criteria for which currently available information is insufficient to indicate a specific recommendation for or against liver transplantation or combined liver and kidney transplantation versus isolated kidney transplantation

C3 mutation^a

CFB mutation^a

Combined mutations among *MCP*, *CFI*, and *CFH*^b

CFHR1-CFHR3 deletion without anti-FH autoantibody^b

Low-risk *CFH* or *CFI* mutation as defined by mutations already reported to the registry as not being associated with recurrence after isolated renal transplantation^b

^aCurrent knowledge is insufficient to presume liver transplantation would correct complement regulation.

^bEvidence is sufficient to support liver or combined liver-kidney transplantation for such individuals if they or a family member with the same mutation experienced aHUS recurrence after isolated kidney transplantation.

Eligibility for isolated kidney transplantation

No evidence of *CFH*, *CFI*, *CFB*, or *C3* gene mutations

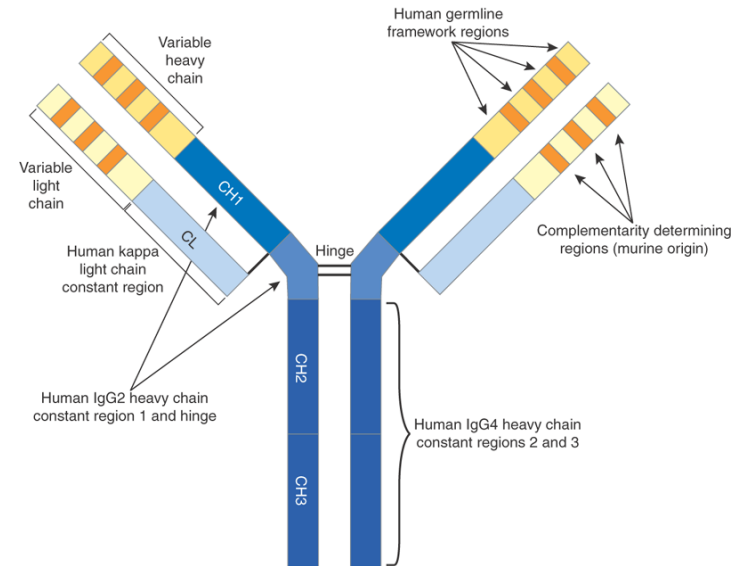
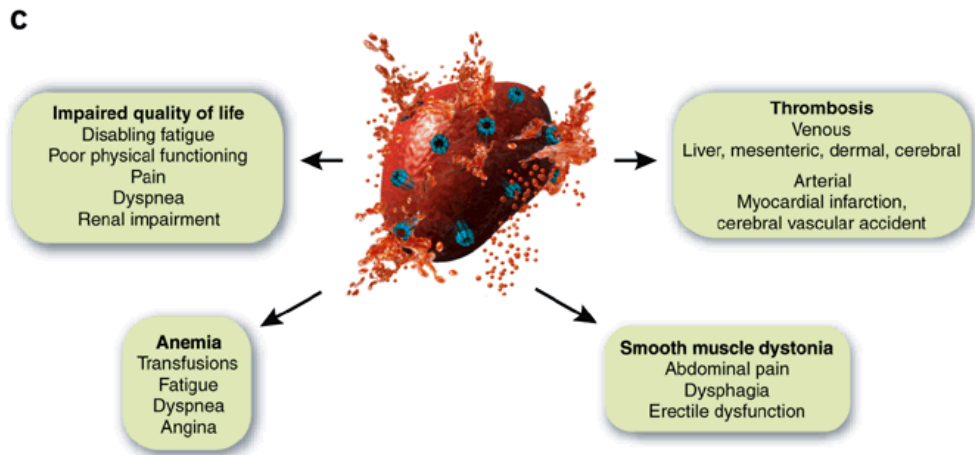
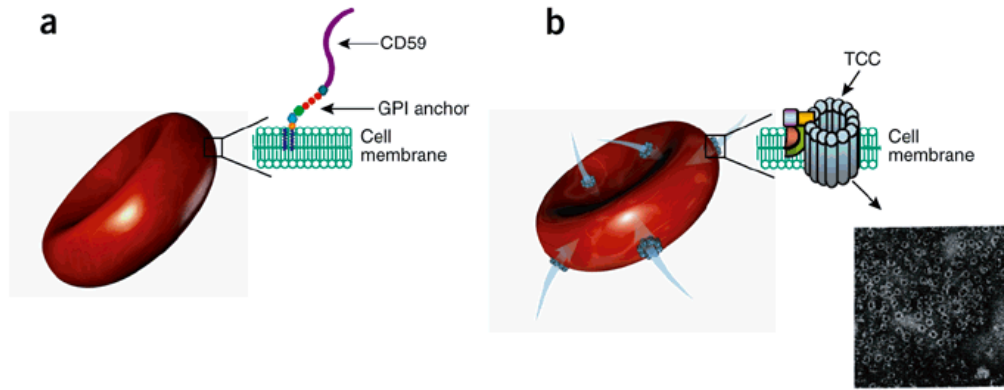
MCP mutation

Mutations associated with successful isolated kidney transplantation in affected family members^a

Anti-factor H autoantibodies

^aConversely, combined liver and kidney transplantation is recommended (see Table 5) for patients who have certain gene mutations and (1) have aHUS recurrence after isolated kidney transplantation or (2) have a family member who had the same mutation and had aHUS recurrence after isolated kidney transplantation.

Eculizumab for paroxysmal nocturnal hemoglobinuria



Eculizumab for aHUS

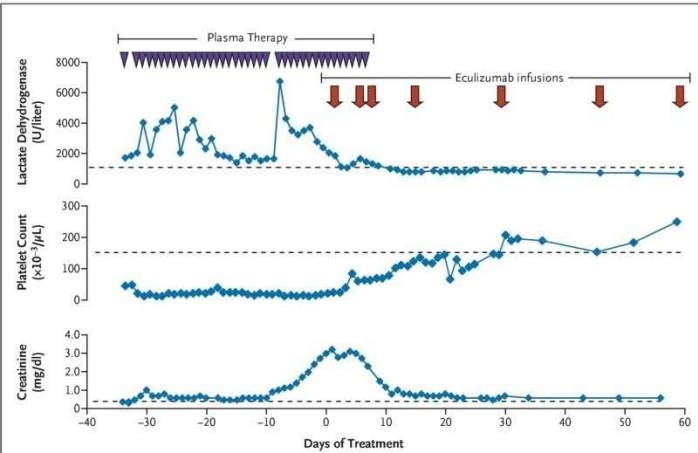


Figure 1. Response to Eculizumab Therapy in a Patient with Congenital Atypical Hemolytic-Uremic Syndrome.
 Eculizumab therapy was initiated in an infant with atypical hemolytic-uremic syndrome for the treatment of a thrombotic microangiopathic event that was unresponsive to 32 consecutive days of plasmapheresis. During the period before treatment with eculizumab, lactate dehydrogenase levels (as a measure of hemolysis) ranged from 1400 to 6800 U per liter (upper limit of the normal range, 920 U per liter [dashed line]) and platelet counts ranged from 11,000 to 38,000 per microliter (lower limit of the normal range, 135,000 per microliter [dashed line]). Creatinine levels began to increase 10 days before the initiation of eculizumab treatment and reached a level of 3.0 mg per deciliter (upper limit of the normal range, 0.4 mg per deciliter [dashed line]). Shortly after eculizumab therapy was initiated, hemolysis decreased (as assessed by levels of lactate dehydrogenase), platelet counts increased, and creatinine levels decreased. Within 1 week after initiation of eculizumab treatment, plasma therapy was discontinued. By day 20, the level of lactate dehydrogenase and the platelet count were normal, and the creatinine level was near normal (0.6 mg per deciliter). Clinical remission was maintained through day 60 with ongoing eculizumab treatment.

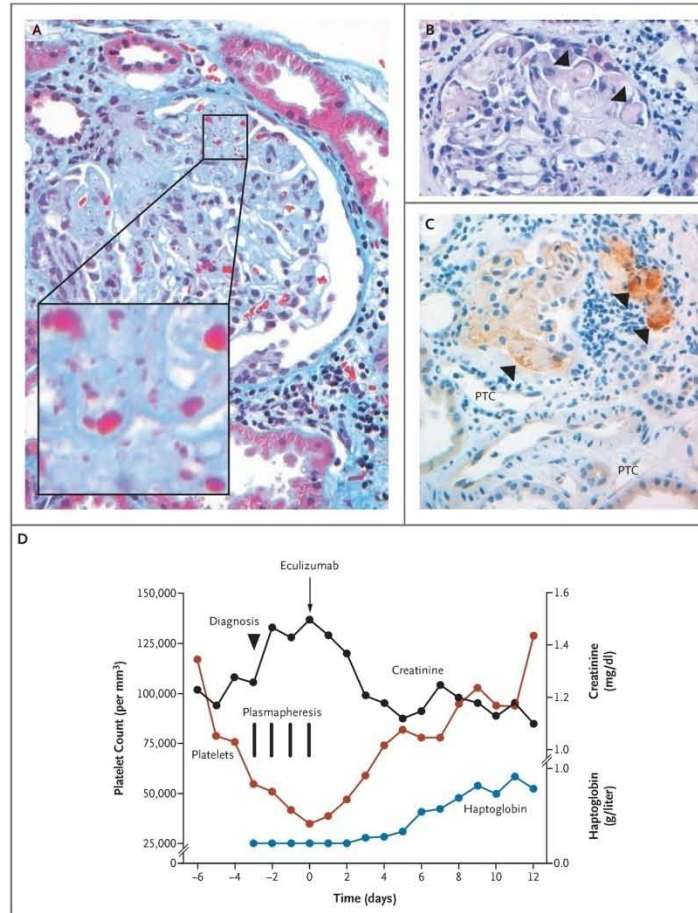
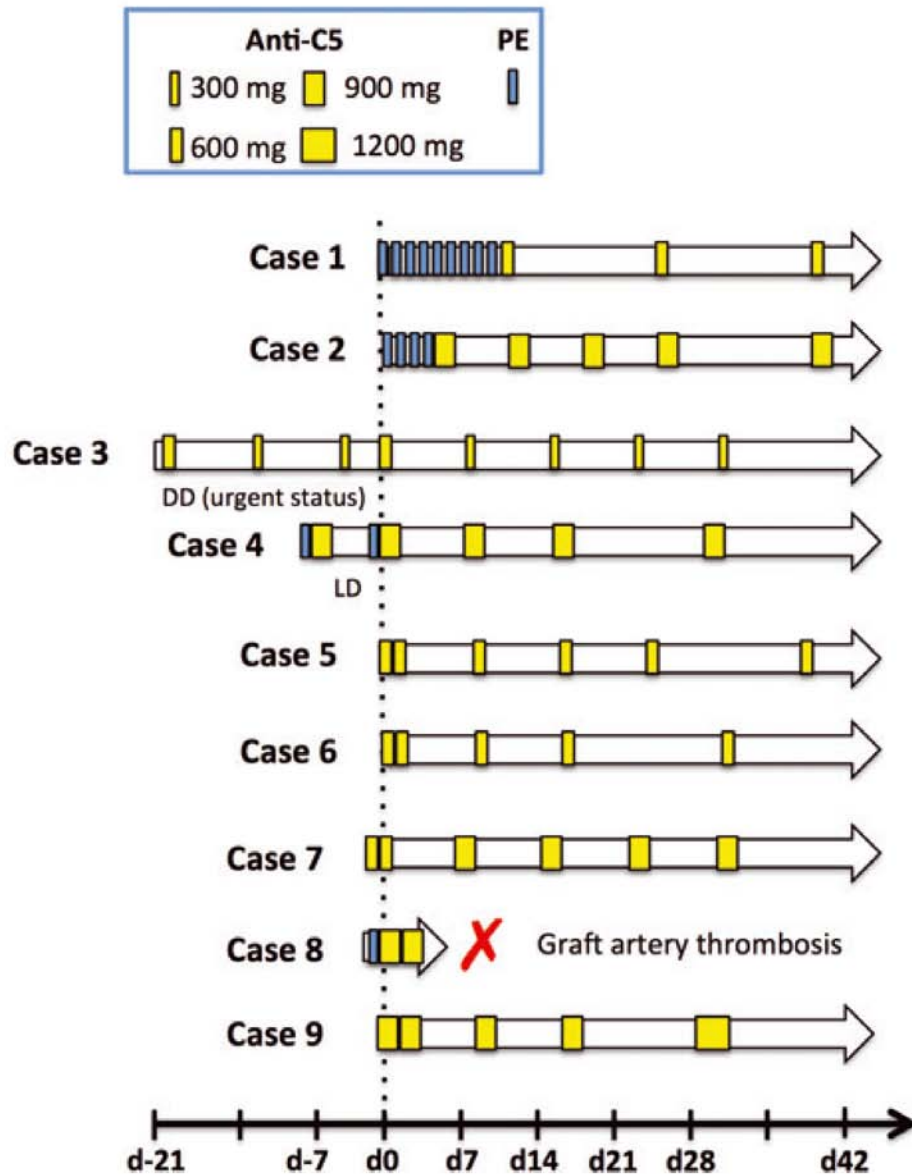


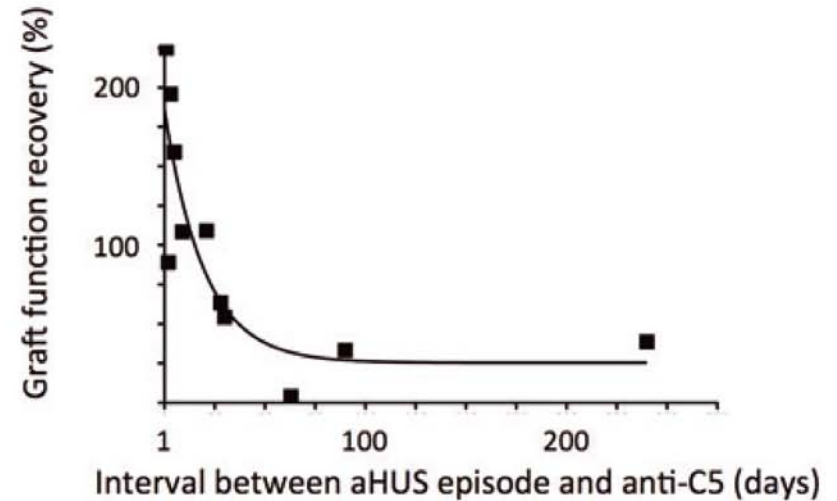
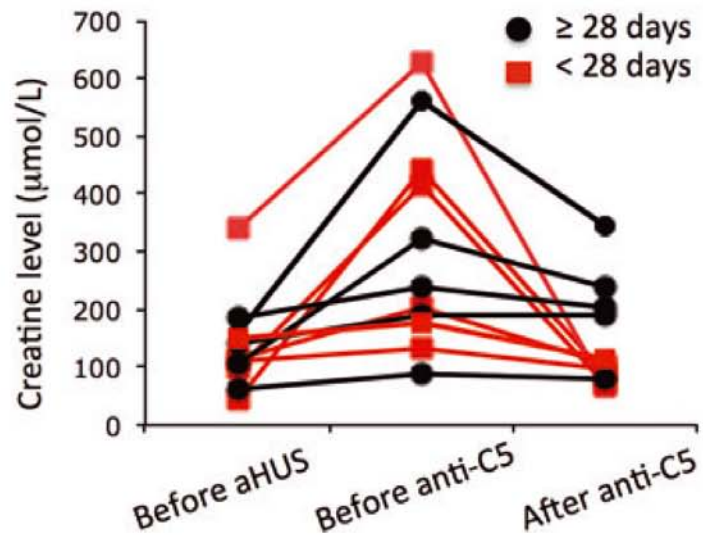
Figure 1 (facing page). Pathological Findings and Laboratory Values.
 In Panel A, elastic Masson's trichrome stain shows thrombotic microangiopathy with numerous intracapillary erythrocytes. The inset shows magnification of the area with red-cell fragments. In Panel B, hematoxylin and eosin stain shows hyaline microthrombi that occlude glomerular capillaries (arrowheads). In Panel C, immunostaining for complement protein C4d was positive in glomerular capillaries and preglomerular arterioles (arrowheads), indicating an activation of the complement system. C4d staining of peritubular capillaries (PTC), an indicator of humoral rejection, was negative. Panel D shows the time course of the platelet count and laboratory values used to determine the occurrence of hemolysis in the patient who underwent renal transplantation (eculizumab was administered at 0 days). When atypical hemolytic-uremic syndrome was diagnosed (arrowhead), tacrolimus was discontinued and plasma exchange was performed four times (vertical black bars indicate treatments). Despite this treatment, atypical hemolytic-uremic syndrome was aggravated, as indicated by the progressive decrease in platelets to 35,000 per cubic millimeter and the deterioration of transplant function indicated by an increase in the creatinine level to 1.5 mg per deciliter. A single dose of 600 mg of eculizumab was administered (arrow), and measurements of total complement activity (data not shown) showed that eculizumab completely blocked the complement system. After treatment with eculizumab, haptoglobin levels normalized after 8 days, the platelet count increased, and the creatinine level decreased, indicating recovery of transplant function.

Prophylactic anti-C5 therapy



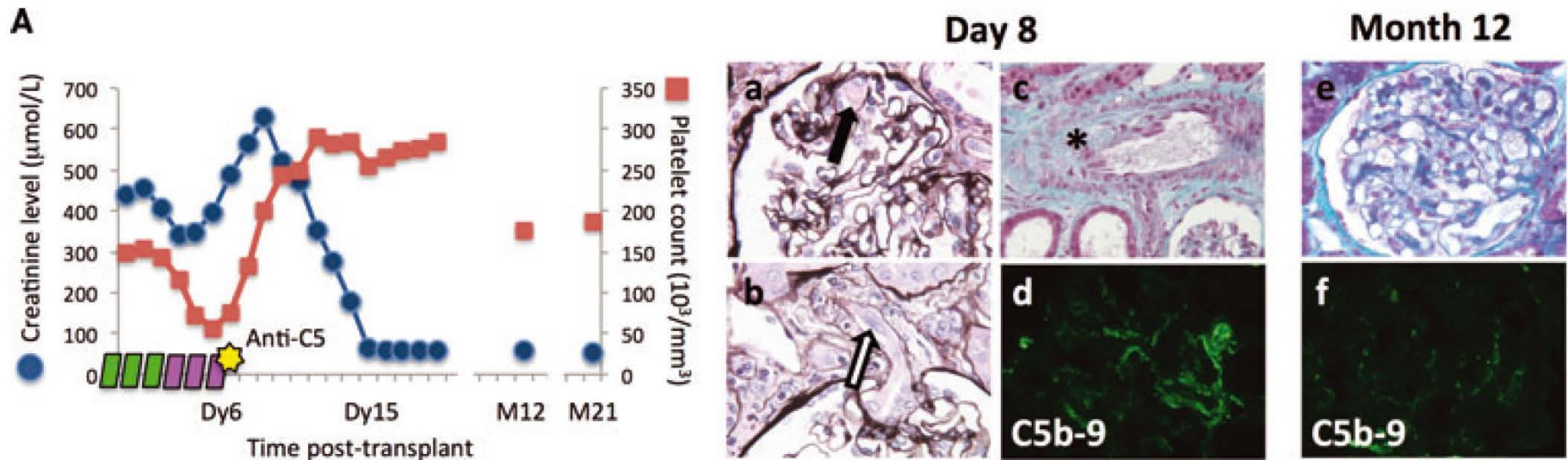
Anti-C5 therapy for post transplant recurrence

Early initiation of anti-C5 therapy is associated with better recovery of renal function.



Anti-C5 therapy for post transplant recurrence

Early initiation of anti-C5 therapy is associated with better recovery of renal function.



What we have learned in the past years

- ❑ Posttransplant aHUS recurrence is a strong predictor of early graft failure
- ❑ Anti-C5 efficiently prevents aHUS posttransplant recurrences
- ❑ Prophylactic plasma therapy may fail to prevent aHUS recurrences
- ❑ Prophylactic plasma therapy may mask subclinical, yet progressive, aHUS recurrences
- ❑ With the advent of anti-C5 therapy, combined kidney-liver transplantation is no longer a first line treatment for aHUS

High risk of recurrence

- ❑ recurrence in a previous graft
mutations associated with a risk of recurrence > 80%
(CFH, CFH-CFHR1 hybrid gene, gain of function mutations in C3, CFB)
- ❑ **prophylactic therapy** (1 additional dose on day 1 post-tx)

Low risk of recurrence

- ❑ mutations in MCP and anti-CFH antibodies
- ❑ treatment if recurrence

Moderate risk of recurrence

- All other patients
- prophylactic therapy (1 additional dose on day 1 post-tx)
- attempt to discontinue treatment 1 year after transplantation

If post-transplant recurrence...

- Prompt initiation of anti-C5 treatment
- Same treatment for full-blown and incomplete aHUS recurrence
- No discontinuation of anti-C5 treatment

aHUS in patients with STEC infections

Two Patients With History of STEC-HUS, Posttransplant Recurrence and Complement Gene Mutations

Alberti et al, Am J Transplant 2013

- ❑ Two patients with a clinical history of STEC-HUS that lost their graft for HUS recurrence
- ❑ Heterozygous CFI mutation in patient #1
Heterozygous MCP mutation in patient #2 , and in her mother who donated the kidney
- ❑ aHUS triggered by STEC infection on a genetic background of impaired complement regulation
- ❑ Complement gene sequencing should be performed before kidney transplantation in patients who developed ESRD following STEC-HUS
- ❑ Genetic analysis of donors is mandatory before living-related transplantation to exclude carriers of HUS-predisposing mutations